

Clarification of the identity of oysters in the genera *Magallana* and *Saccostrea* in the upper Gulf of Thailand based on 16S rRNA sequences

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Abstract. Oysters are ecologically important, often dominating shorelines and becoming habitat modifiers. They are a key group for molluscan aquaculture, with an estimated annual production of 8,000 tonnes in Thailand. Some species become invasive when introduced into new habitats. A variety of species have been recorded from Thailand, but identifications are uncertain. We examined oysters at 14 sites in the upper (northern) Gulf of Thailand in August 2023 and compared identifications made in the field using Thai literature with DNA sequences. Two species of *Magallana* and four species of *Saccostrea* were detected in the wild, and one species of each genus in aquaculture farms. The taxonomy of the species was clarified and their invasive potential evaluated.

Keywords. *Saccostrea*, *Magallana*, *Crassostrea*, invasive marine species, marine pests, Ostreidae, competition, bivalves

INTRODUCTION

Global aquaculture production has increased dramatically in recent decades and now contributes to more than 50% of the world supply of seafood. Molluscs were the second largest component of total production at 21% both by weight and value in 2016, with oysters dominating molluscan aquaculture production. Many oyster species are suitable for aquaculture as they are easily cultured, fast growing, feed naturally, and are commercially valuable. The total annual world aquaculture production is about six million tonnes, dominated by *Magallana gigas* (Thunberg, 1793). China is by far the largest producer, contributing 86% of the total global production, but very little is exported. A number of other countries such as France, United States of America, South Korea, Japan, and Canada have larger export markets (Botta et al., 2020).

Oyster aquaculture is the third most important shellfish culture in Thailand. The Thai Department of Fisheries reported a total estimated production of about 8,000 tonnes from 1,000 farms with a total area of 9.6 km² in 2022. Nearly 600 farms in

eastern Thailand contribute about 85% of the total production (DoF, 2023). The key commercial species were listed as *Saccostrea commercialis* (Iredale & Roughley, 1933), *Crassostrea lugubris* (Sowerby II, 1871), and *Crassostrea belcheri* (Sowerby II, 1871) (Trivej & Kesjinda, 2018). Jearnsripong et al. (2022) estimated that oyster production in the Gulf of Thailand in 2021 was 13,101 tonnes with a value of THB 255.4 million; in contrast, production from the Andaman Sea coast was only 216 tonnes worth THB 24.5 million. The importance of oyster aquaculture to farming communities at Ang Sila in the upper Gulf of Thailand has been highlighted by Szuster et al. (2008).

The densely packed aggregations of many oysters cause considerable variations in shell form, making accurate taxonomic identification very difficult. Bussarawit & Cedhagen (2010) reported that previous studies on oysters in Thailand were scarce and there was considerable confusion regarding the scientific names with inconsistent taxonomic classification, making identifications of species reported in most published information uncertain. In some cases, unidentified or misidentified oysters were preserved in the collection of the Phuket Marine Biological Center (Nielsen, 1976; Tantanawong, 1979); these were subsequently revised by Bussarawit & Cedhagen (2010). DNA analysis is the most reliable method for identifying species (Salvi & Mariottini, 2016; Willan et al., 2021). Bussarawit et al. (2006) and Klinbunga et al. (2000, 2001, 2002, 2003, 2005) studied the genetics of several species of Thai oysters but the taxonomy they used, primarily based on Harry (1985), is now outdated. There have been major changes to ostreid taxonomy in recent years (e.g., Huber, 2010; Salvi & Mariottini, 2016; Sekino & Yamashita, 2016; Sigwart et al., 2021). Bussarawit et al. (2006) used partial 16S rRNA sequences to show that specimens identified as *Saccostrea*

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forskahlii (Gmelin, 1791) and *Saccostrea* cf. *malabonensis* (Faustino, 1932) were genetically identical despite obvious morphological differences. Their results also indicated that a specimen identified as *Striostrea mytiloides* (Lamarck, 1819) belonged to the genus *Saccostrea*. Accordingly, the first goal of this paper is to use DNA sequencing to determine the identity of oysters in the genera *Magallana* and *Saccostrea* in the upper Gulf of Thailand and to, as far as possible, reconcile the current taxonomy with published records for the Gulf of Thailand.

In recent years, there has been increasing concern over the threat to global marine environments posed by invasive marine species (IMS, also known as introduced marine pests) (e.g., Johnson & Chapman, 2007; Molnar et al., 2008; Katsanevakis et al., 2014; Crowe & Frid, 2015; McDonald et al., 2020; Salimi et al., 2021). Unfortunately, the biological attributes that make oysters attractive for aquaculture also increase their IMS potential. Australian authorities have developed strong quarantine barriers to prevent the arrival of IMS to Australian waters (e.g., Vessel-Check, 2022) and monitoring strategies for species that penetrated the barriers (NIMPCG, 2010a, b; DPIRD, 2016). The introduction of oysters for aquaculture also brings risks of introducing diseases and other species living on or in the oysters (Ruesink et al., 2005; Herbert et al., 2016). The second goal of this paper is to determine what species of *Magallana* and *Saccostrea* are in fact present in the Gulf of Thailand and to assess their IMS risk to Australia.

MATERIAL AND METHODS

Sample collection. Oysters in the genera *Magallana* and *Saccostrea* were surveyed at 14 sites in the upper Gulf of Thailand in August 2023, in conjunction with a survey for the invasive mytilid *Mytella strigata* (Hanley, 1843) (Wells et al., 2024). The oyster survey commenced at Sriracha in Chonburi Province on the eastern side of the upper gulf on 1 August (Fig. 1; Table 1) and proceeded south to Kungkraben Bay. At most sites intertidal rocky shores were sampled on low spring tides. At Sriracha a small boat was used to inspect four commercial Asian green mussel aquaculture lines. At Kungkraben Bay a rocky shore was sampled near the base of the aquaculture demonstration facility, where hanging ropes in the facility were also inspected. Sites on the western side of the inner Gulf of Thailand were then examined, beginning at Klong Wan, then moving north back to Bangkok. The survey was completed on 8 August 2023.

At each site oysters were identified in the field as far as possible using Bussarawit & Cedhagen (2010) (see Table 2). Specimens of each presumed species and any doubtful species were removed by hand from the rocks using a hammer and chisel and were individually labelled. The oysters were opened, a small tissue sample was dissected from the adductor muscle, and preserved in 95% ethanol. The remaining tissue was discarded. Each oyster shell was then placed in a labelled plastic bag with bleach to remove any attached tissue.

Oyster specimens were collected in four markets in Chonburi in March 2024 (Fig. 1). These oysters were taken to the Kasetsart University Museum of Fisheries (Natural History) (KUMF) in Bangkok and were handled as described below.

Full details, including the museum and GenBank registration numbers of all specimens used in this study, are presented in Table 1.

Taxonomy. At the KUMF, the oyster shells were washed in freshwater, cleaned with a wire brush, dried, and photographed. Most specimens were deposited in the KUMF. Voucher representatives of each species were hand carried to Singapore and deposited in the Lee Kong Chian Natural History Museum (LKCNHM) of the National University of Singapore. Taxonomy in this paper follows the World Register of Marine Species (WoRMS) (MolluscaBase eds., 2024). Published literature was searched and all Gulf of Thailand records examined.

Other abbreviations are: ANG, Ang Sila; BAN, Bang Saen; PCR, Polymerase Chain Reaction; ZRC, Zoological Reference Collection, Lee Kong Chian Natural History Museum, National University of Singapore; RV, right valve; LV, left valve.

DNA extraction, PCR amplification, and sequencing.

Tissue samples were prepared for each species identified at each site. The adductor muscle was dissected with small pieces of tissue being removed and preserved in 95% ethanol in a small vial for DNA analysis. A small piece of absorbent cloth was inserted into each vial, and the vial drained and sealed. The vials were packed and hand carried with the necessary documentation to a laboratory at Curtin University in Perth. At Curtin the vials were frozen until analysis could be undertaken. For sequencing, the tissue from each vial was thawed and DNA extracted using the DNeasy Blood and Tissue Kit (Qiagen Inc., USA) following the manufacturer's instructions. PCR amplification of approximately 450 bases of the mitochondrial ribosomal RNA subunit 16S region (16S) was done for each specimen, using primers 16Sar and 16Sbr (Simon et al., 1994).

PCR reactions were conducted in 25 µl containing 3 µl DNA (~20 ng), 1× Invitrogen Platinum Green Hot Start PCR master mix (containing 1.5 mM MgCl₂ and 0.2mM of each dNTP), 0.5 mg/ml bovine serum albumin (Fisher Biotec, Australia), and 0.6 µM of each primer. PCR conditions consisted of an initial incubation at 95°C for 3 minutes, followed by 35 cycles of 94°C for 45 seconds, 52°C for 90 seconds, 72°C for 45 seconds, and a final extension step of 72°C for 10 minutes.

Bi-directional sequencing of unpurified PCR products was performed using the Sanger sequencing service provided by the Australian Genome Research Facility, Perth. Sequences were trimmed and edited using the Geneious Prime 2022.1.1 software (<http://www.geneious.com>). For each individual, species identification was verified by similarity-based searches on the NCBI BLAST database (Altschul et al.,



Fig. 1. Map of the upper (northern) Gulf of Thailand showing the locations of stations sampled in August 2023 and the aquaculture farms in Chonburi. ANG, Ang Sila; BAN, Bang Saen. Details of the locations are provided in Table 1.

1990). All sequences were submitted to GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>) and assigned individual accession numbers.

Additional 16S sequences for East Asian *Magallana* and *Saccostrea* species were retrieved from GenBank and aligned using the Geneious Prime 2022.1.1 software. Sequences that were not within the targeted region of the 16S rRNA gene or were very short were removed. jModelTest v2.1.10 (Darriba et al., 2012) was used to find the best evolutionary model of nucleotide substitution for the alignments. Specimens identified as *Magallana* species were aligned with *Magallana* sequences retrieved from GenBank and this alignment was trimmed to 418 bp. For this alignment we performed a Bayesian Inference analysis using the MrBayes v3.2.6 (Huelsenbeck & Ronquist, 2001) plugin in Geneious Prime with the following parameters: HKY85 gamma model with nucleotide sites partitioned for 1 million generations subsampling every 500 generations and a 100,000 burn-in length. Likewise, all remaining sequences were aligned with *Saccostrea* sequences retrieved from GenBank and this alignment was trimmed to 435 bp. For

this alignment, we performed a Bayesian Inference analysis using the MrBayes v3.2.6 (Huelsenbeck & Ronquist, 2001) plugin in Geneious Prime with the following parameters: HKY85 invgamma model with nucleotide sites partitioned for 1 million generations subsampling every 500 generations and a 100,000 burn-in length. For both Bayesian inference analyses, *Ostrea edulis* Linnaeus, 1758 was used as an outgroup. All resultant trees were processed in FigTree v. 1.4.4 (Rambaut et al., 2018) with tree annotations added in Adobe Illustrator.

Full details of all sequences used in this study are included in Table 1 and Appendix 1.

TAXONOMY

Eighty-nine oysters were collected from 14 stations in the upper Gulf of Thailand in August 2023; partial 16S rRNA sequences were obtained from 67 oysters. Additionally, 46 oysters from farms in Chonburi were bought in four markets; 33 of these were sequenced.

Table 1. Site locations and specimens used in this study. The sequence identification codes (SIC) correspond to those used in Figs. 2 and 3.

Site no.	Location	Latitude and longitude	Date	Habitat	Bussarawit & Cedhagen (2010)	Species detected			
						Species	SIC	Specimen voucher number	GenBank accession number
1	Kasetsart University Fisheries Research Center, Sriracha	13°10.728'N; 100°55.258'E	1 August 2023	Hanging ropes	<i>C. bilineata</i> , <i>S. cucullata</i>	<i>M. bilineata</i>	TH01	KUMF-MOLL 1207	PP836532
						<i>M. bilineata</i>	TH02	ZRC.MOL.028369	PP836533
						<i>M. bilineata</i>	TH03	ZRC.MOL.028370	PP836534
						<i>Saccostrea</i> F	TH08	ZRC.MOL.028371	PP836535
						<i>Saccostrea</i> F	TH10	KUMF-MOLL 1213	PP836536
						<i>M. belcheri</i>	TH26	ZRC.MOL.028372	PP836537
						<i>M. belcheri</i>	TH27	KUMF-MOLL 1214	PP836538
						<i>M. belcheri</i>	TH28	KUMF-MOLL 1215	PP836539
						<i>S. spathulata</i>	TH35	KUMF-MOLL 1216	PP836540
						<i>S. spathulata</i>	TH36	KUMF-MOLL 1217	PP836541
2	Samaesan and Seafood market	12°35.955'N; 100°56.948'E	2 August 2023	Rocky shore	<i>S. cucullata</i> , <i>S. echinata</i> , <i>Ostrea</i> sp., <i>C. belcheri</i> in market	<i>S. spathulata</i>	TH37	KUMF-MOLL 1218	PP836542
						<i>S. spathulata</i>	TH38	ZRC.MOL.028373	PP836543
						<i>S. spathulata</i>	TH39	ZRC.MOL.028374	PP836544
						<i>S. spathulata</i>	TH41	KUMF-MOLL 1220	PP836545
						<i>S. spathulata</i>	TH42	ZRC.MOL.028375	PP836546
						<i>S. scyphophilla</i>	TH43	KUMF-MOLL 1221	PP836547
						<i>S. scyphophilla</i>	TH44	KUMF-MOLL 1222	PP836548
						<i>S. scyphophilla</i>	TH45	–	PP836549
						<i>Saccostrea</i> B	TH46	ZRC.MOL.028376	PP836550
						<i>S. scyphophilla</i>	TH48	KUMF-MOLL 1224	PP836551
3	Ta Kuan Beach	12°40.295'N; 101°09.887'E	2 August 2023	Rocky shore	<i>S. forskahlii</i>	<i>Saccostrea</i> B	TH49	ZRC.MOL.028377	PP836552
						<i>Saccostrea</i> B	TH58	ZRC.MOL.028378	PP836553
						<i>Saccostrea</i> B	TH59	KUMF-MOLL 1226	PP836554
						<i>Saccostrea</i> F	TH61	–	PP836555
4	Mae Rum Peung Beach	12°36.638'N; 101°23.028'E	2 August 2023	Rocky shore	<i>Striosorea</i> sp., <i>S. cucullata</i> , <i>S. echinata</i>	<i>Saccostrea</i> B	TH65	KUMF-MOLL 1228	PP836556
						<i>Saccostrea</i> B	TH66	KUMF-MOLL 1229	PP836557
						<i>Saccostrea</i> B	TH68	KUMF-MOLL 1231	PP836558
5	Hin Khrong north of Kungkraben Bay	12°36.475'N; 101°52.205'E	3 August 2023	Rocky shore	<i>S. cucullata</i> , <i>Striosorea</i> sp.	<i>Saccostrea</i> B	TH69	KUMF-MOLL 1232	PP836559
						<i>Saccostrea</i> B	TH72	KUMF-MOLL 1235	PP836560
						<i>Saccostrea</i> B	TH74	KUMF-MOLL 1237	PP836561

Site no.	Location	Latitude and longitude	Date	Habitat	Bussarawit & Cedhagen (2010)	Species detected				
						Species	SIC	DNA sequence		
								Specimen voucher number	GenBank accession number	
6	Mouth of Kungkraben Bay	12°35.213'N; 101°53.062'E	3 August 2023	Rocky shore	<i>S. echinata</i> , <i>S. forskahlii</i> , <i>C. bilineata</i>	<i>M. bilineata</i>	TH88	KUMF-MOLL	1238	PP836562
						<i>M. bilineata</i>	TH90	KUMF-MOLL	1240	PP836563
						<i>Saccostrea</i> B	TH92	KUMF-MOLL	1242	PP836564
						<i>Saccostrea</i> B	TH93	-		PP836565
						<i>Saccostrea</i> F	TH94	KUMF-MOLL	1243	PP836566
						<i>Saccostrea</i> F	TH95	ZRC.MOL.028379		PP836567
7	Kungkraben Bay adjacent to Sea Farming Demonstration Unit	12°34.898'N; 101°53.507'E	3 August 2023	Rocky shore and wall	<i>C. bilineata</i> , <i>S. cucullata</i> , <i>S. forskahlii</i> , <i>Striostrea</i> sp.	<i>M. bilineata</i>	TH98	KUMF-MOLL	1246	PP836568
						<i>M. bilineata</i>	TH99	KUMF-MOLL	1247	PP836569
						<i>Saccostrea</i> B	TH102	ZRC.MOL.028380		PP836570
						<i>Saccostrea</i> B	TH105	ZRC.MOL.028381		PP836571
						<i>Saccostrea</i> F	TH106	ZRC.MOL.028382		PP836572
						<i>Saccostrea</i> B	TH108	KUMF-MOLL	1253	PP836573
8	Kungkraben Bay, Sea Farming Demonstration Unit	12°34.898'N; 101°53.507'E	3 August 2023	Hanging ropes	<i>S. cucullata</i>	None collected as it is a research area.	TH109	KUMF-MOLL	1254	PP836574
9	Pattaya	12°58.422'N; 100°53.373'E	4 August 2023	Rocky shore and wall	<i>C. bilineata</i> , <i>S. cucullata</i> , <i>S. forskahlii</i> , <i>Striostrea</i> sp.	<i>M. bilineata</i>	TH117	KUMF-MOLL	1258	PP836575
						<i>M. bilineata</i>	TH118	KUMF-MOLL	1259	PP836576
						<i>Saccostrea</i> F	TH121	ZRC.MOL.028383		PP836577
						<i>Saccostrea</i> B	TH123	KUMF-MOLL	1263	PP836578
						<i>Saccostrea</i> B	TH126	ZRC.MOL.028384		PP836579
						<i>Saccostrea</i> B	TH128	KUMF-MOLL	1267	PP836580
10	Klong Wan Pier	11°44.133'N; 99°46.915'E	6 August 2023	Jetty	<i>S. forskahlii</i> , <i>Striostrea</i> sp.	<i>Saccostrea</i> F	TH130	KUMF-MOLL	1269	PP836581
						<i>Saccostrea</i> B	TH132	KUMF-MOLL	1271	PP836582
						<i>Saccostrea</i> F	TH134	KUMF-MOLL	1273	PP836583
						<i>Saccostrea</i> B	TH137	KUMF-MOLL	1276	PP836584
11	Khao Ta Mong Lai Forest Park	11°50.198'N; 99°49.807'E	6 August 2023	Rocky shore	<i>S. forskahlii</i> , <i>S. cucullata</i>	<i>Saccostrea</i> F	TH139	KUMF-MOLL	1278	PP836585
						<i>Saccostrea</i> B	TH140	KUMF-MOLL	1279	PP836586
						None collected as it is a national park.				

Site no.	Location	Latitude and longitude	Date	Habitat	Bussarawit & Cedhagen (2010)	Species detected			
						Species	SIC	DNA sequence	
									Specimen voucher number
12	Sirinart Rajini Mangrove Ecology Learning Center	12°23.573'N; 99°58.944'E	7 August 2023	<i>Rhizophora</i> mangroves	<i>Striostrea</i> sp., <i>S. forskahlii</i>	None collected as it is a national park.			
13	Khao Ta Kiab Pier	12°30.812'N; 99°58.840'E	7 August 2023	Jetty	<i>Striostrea</i> sp., <i>S. forskahlii</i> , <i>S. cucullata</i>	<i>Saccostrea</i> F	THI41	KUMF-MOLL 1280	PP836587
						<i>Saccostrea</i> F	THI42	KUMF-MOLL 1281	PP836588
						<i>Saccostrea</i> B	THI43	KUMF-MOLL 1282	PP836589
						<i>Saccostrea</i> B	THI45	KUMF-MOLL 1284	PP836590
						<i>Saccostrea</i> F	THI46	KUMF-MOLL 1285	PP836591
						<i>Saccostrea</i> F	THI47	KUMF-MOLL 1286	PP836592
						<i>Saccostrea</i> B	THI48	KUMF-MOLL 1287	PP836593
						<i>Saccostrea</i> F	THI49	KUMF-MOLL 1288	PP836594
14	Institute of Marine Science, Burapha University (Cha Am Substation)	12°52.283'N; 100°01.067'E	8 August 2023	Rock wall	<i>Striostrea</i> sp., <i>S. forskahlii</i>	<i>Saccostrea</i> B	THI52	KUMF-MOLL 1291	PP836595
						<i>Saccostrea</i> B	THI53	KUMF-MOLL 1292	PP836596
						<i>Saccostrea</i> B	THI55	KUMF-MOLL 1294	PP836597
						<i>Saccostrea</i> B	THI56	KUMF-MOLL 1295	PP836598
Market 1	Chonburi market	13°18.664'N; 100°54.784'E	1 March 2023		<i>C. bilineata</i> ; <i>S. forskahlii</i>	<i>Saccostrea</i> F	THI57	KUMF-MOLL 1306	PP836599
						<i>M. bilineata</i>	THI58	KUMF-MOLL 1304	PP836600
						<i>M. bilineata</i>	THI59	KUMF-MOLL 1303	PP836601
						<i>Saccostrea</i> F	THI60	KUMF-MOLL 1297	PP836602
						<i>Saccostrea</i> F	THI61	KUMF-MOLL 1299	PP836603
						<i>Saccostrea</i> F	THI62	KUMF-MOLL 1302	PP836604
						<i>Saccostrea</i> F	THI85	KUMF-MOLL 1300	PP836627
						<i>Saccostrea</i> F	THI86	KUMF-MOLL 1305	PP836628
Market 2	Chonburi market	13°18.830'N; 100°55.003'E	1 March 2023		<i>C. bilineata</i> ; <i>S. forskahlii</i>	<i>Saccostrea</i> F	THI63	KUMF-MOLL 1308	PP836605
						<i>Saccostrea</i> F	THI64	KUMF-MOLL 1310	PP836606
						<i>Saccostrea</i> F	THI65	KUMF-MOLL 1309	PP836607
						<i>M. bilineata</i>	THI66	KUMF-MOLL 1314	PP836608
						<i>M. bilineata</i>	THI67	KUMF-MOLL 1316	PP836609
						<i>M. bilineata</i>	THI68	KUMF-MOLL 1318	PP836610
						<i>Saccostrea</i> F	THI81	KUMF-MOLL 1313	PP836623
						<i>M. bilineata</i>	THI82	KUMF-MOLL 1315	PP836624
						<i>M. bilineata</i>	THI89	KUMF-MOLL 1317	PP836631

Site no.	Location	Latitude and longitude	Date	Habitat	Bussarawit & Cedhagen (2010)	Species detected				
						Species	SIC	DNA sequence		
								Specimen voucher number	GenBank accession number	
Market 3	Chonburi market	13°19.003'N; 100°55.122'E	1 March 2023		<i>C. bilineata</i> ; <i>S. forskahlii</i>	<i>Saccostrea</i> F	TH169	KUMF-MOLL	1321	PP836611
						<i>Saccostrea</i> F	TH170	KUMF-MOLL	1322	PP836612
						<i>Saccostrea</i> F	TH171	KUMF-MOLL	1326	PP836613
						<i>M. bilineata</i>	TH172	KUMF-MOLL	1329	PP836614
						<i>M. bilineata</i>	TH173	KUMF-MOLL	1330	PP836615
						<i>M. bilineata</i>	TH174	KUMF-MOLL	1332	PP836616
						<i>Saccostrea</i> F	TH183	KUMF-MOLL	1324	PP836625
						<i>M. bilineata</i>	TH184	KUMF-MOLL	1328	PP836626
						<i>M. bilineata</i>	TH175	KUMF-MOLL	1333	PP836617
						<i>Saccostrea</i> F	TH176	KUMF-MOLL	1335	PP836618
Market 4	Chonburi market	13°19.937'N; 100°55.382'E	1 March 2023		<i>C. bilineata</i> ; <i>S. forskahlii</i>	<i>Saccostrea</i> F	TH177	KUMF-MOLL	1337	PP836619
						<i>M. bilineata</i>	TH178	KUMF-MOLL	1340	PP836620
						<i>M. bilineata</i>	TH179	KUMF-MOLL	1339	PP836621
						<i>M. bilineata</i>	TH180	KUMF-MOLL	1341	PP836622
						<i>M. bilineata</i>	TH187	KUMF-MOLL	1338	PP836629
						<i>Saccostrea</i> F	TH188	KUMF-MOLL	1334	PP836630

Figs. 2 and 3 show the phylogenetic relationships of the *Magallana* and *Saccostrea* species respectively, based on material collected in the upper Gulf of Thailand and other GenBank registrations of species in both genera from Southeast Asia. The European *Ostrea edulis* was used as the outgroup. Two species of *Magallana* were recorded: *Magallana belcheri* (Sowerby II, 1871) and *Magallana bilineata* (Röding, 1798). Four genetic lineages of *Saccostrea* were collected in the wild: *Saccostrea* lineages B and F, *Saccostrea spathulata* (Lamarck, 1819) (= lineage J), and *Saccostrea scyphophilla* (Péron & Lesueur, 1807). Only two species were recorded from the farmed aquaculture specimens: *Magallana bilineata* and *Saccostrea* lineage F.

Bussarawit et al. (2006) sequences were excluded from the Bayesian Inference analysis since they were shorter than the rest of the dataset. However, during the alignment of sequences, it was noted that the sequences assigned to *Saccostrea forskahlii* (EF122388) and *S. cf. malabonensis* (EF122391) were 99.8–100% matches to specimens collected in this study that made up the *Saccostrea* lineage F clade (Fig. 3). *Saccostrea cucullata* (Born, 1778) (EF122389) sequence was 99.4–100% match to specimens that are part of the *Saccostrea scyphophilla* clade (Fig. 3) and *Saccostrea echinata* (Quoy & Gaimard, 1835) (EF122390) sequence was 99.4–100% match to specimens in the *Saccostrea spathulata* (Lamarck, 1819) clade. Even though Bussarawit et al. (2006) had a small sample size, they had similar species/lineage detections in Thailand based on partial 16S rRNA sequences.

Family Ostreidae Rafinesque, 1815

Subfamily Crassostreinae Scarlato & Starobogotov, 1979

Genus *Magallana* Salvi & Mariottini, 2016

Members of the well-known genus *Crassostrea* Sacco, 1897 were defined based on shell morphology and included species from the Pacific, Indian, and Atlantic Oceans. The genus included several commercially and ecologically important species, particularly *Crassostrea gigas* (Thunberg, 1793) in the Pacific and *Crassostrea virginica* (Gmelin, 1791) in the Atlantic. *Crassostrea gigas* was also regarded as an invasive marine species (NIMPCG, 2010a, b; DPIRD, 2016). However, with the development and use of DNA techniques it became clear that there were consistent genetic differences between extant species of *Crassostrea* in the Indo-Pacific and the Atlantic (Ó'Foighil et al., 1995; Lam & Morton, 2004; Wang et al., 2004a, b). Salvi et al. (2014) and Salvi & Mariottini (2016) proposed the separation of *Crassostrea* into three genera: *Crassostrea* sensu

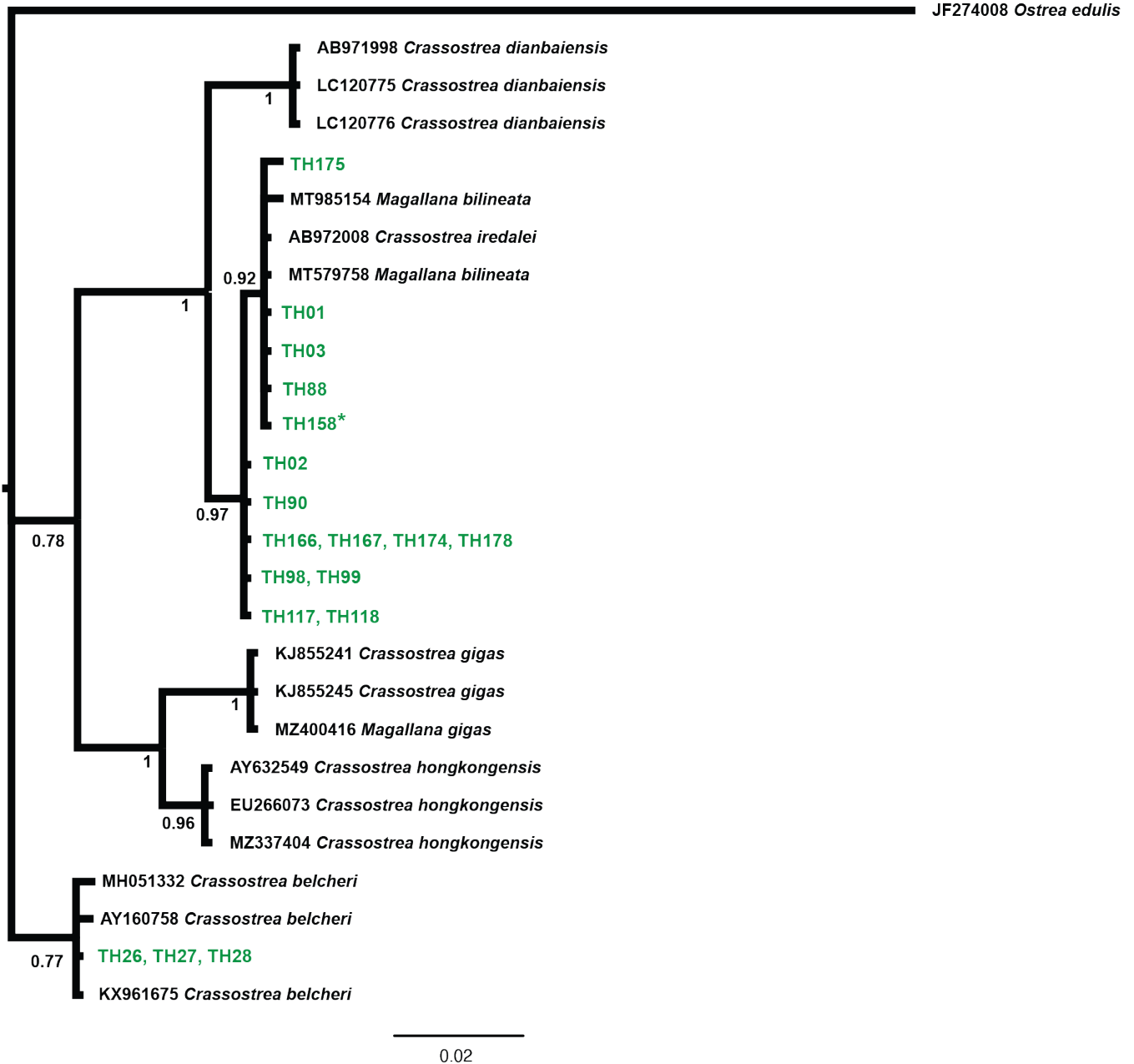


Fig. 2. 16S Bayesian inference tree of Southeast Asian *Magallana* species using *Ostrea edulis* as the outgroup. Genus names shown are those associated with the GenBank accession numbers. Sequence identification codes with prefixes TH in green font represent individuals sequenced in this study. Bootstrap values are shown at the nodes. TH158* (*Magallana bilineata*) represents specimens TH158, TH159, TH168, TH172, TH173, TH179, TH180, TH182, TH184, TH187, and TH189.

stricto for Atlantic species, and *Talonostrea* Li & Qi, 1994 and *Magallana* Salvi & Mariottini, 2016 for Indo-Pacific species. The proposed separation of Atlantic and Indo-Pacific *Crassostrea* has been controversial. Several researchers (Bayne et al., 2017, 2019; Backeljau, 2018) argued for nomenclatural stability as *Crassostrea* is well understood and includes species that are ecologically and economically important. Salvi & Mariottini (2021), Willan (2021) and Salvi et al. (2022) believed the separation of *Crassostrea* should be accepted as the genetic data are clear and there have been numerous nomenclatural changes of economically and ecologically important species in other taxa as new

information became available. Sigwart et al. (2021) observed that there are clearly identifiable extant and fossil species that, in the absence of genetic data, cannot be included in one of the three genera. They suggested retaining the genus *Crassostrea* and recognising three subgenera (*Crassostrea*, *Magallana*, and *Talonostrea*). WoRMS recognises the three as valid genera (MolluscaBase eds., 2024), a classification followed by Tan et al. (2024) for *Magallana* in Singapore. We have followed the WoRMS use of *Magallana* and the WoRMS recognised synonymy of the species.

Magallana species present in the upper Gulf of Thailand:

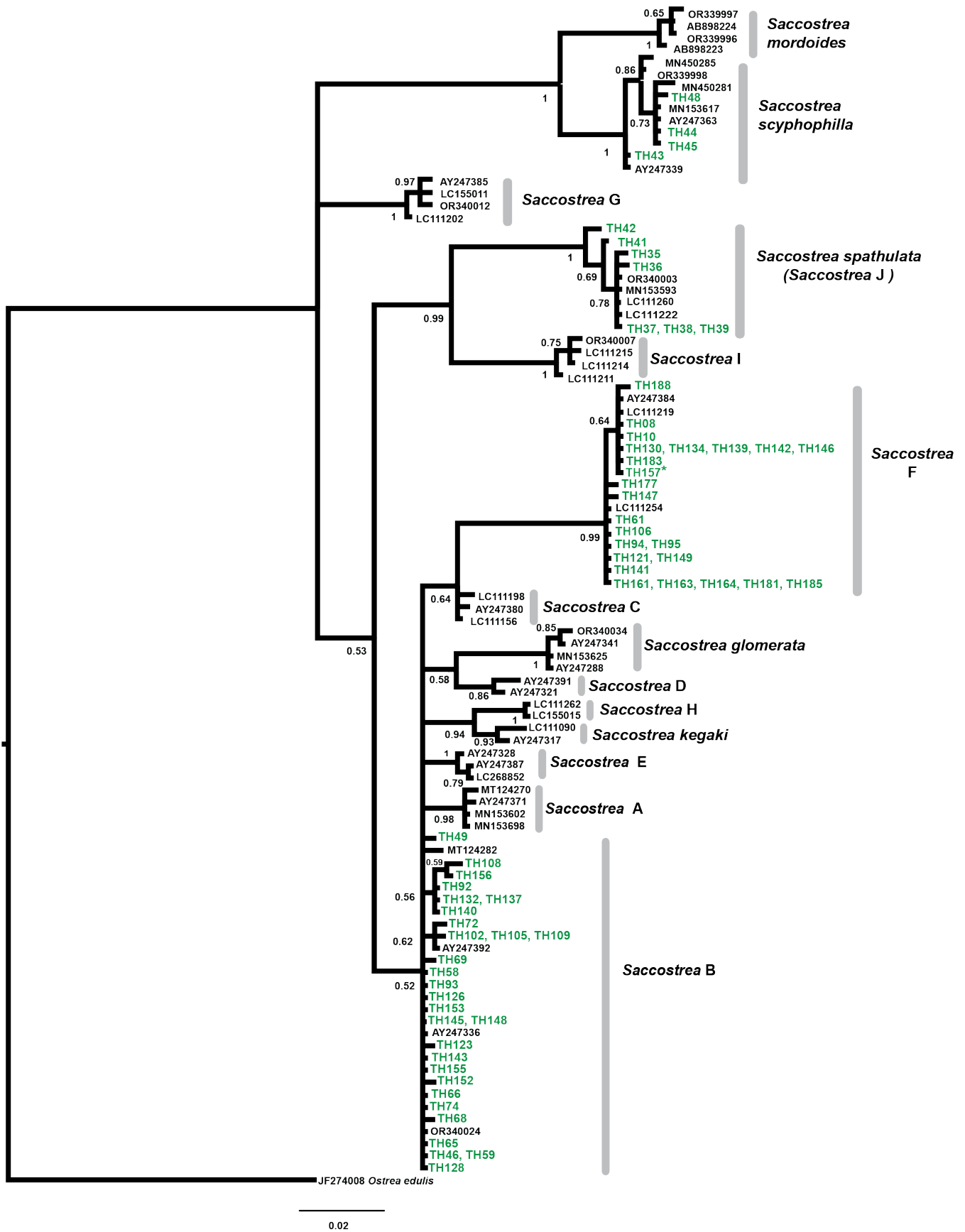


Fig. 3. 16S Bayesian inference tree of Asian *Saccostrea* species using *Ostrea edulis* as the outgroup. Sequence identification codes with prefixes TH in green font represent individuals sequenced in this study. Bootstrap values are shown at the nodes. TH157* (*Saccostrea* lineage F) represents specimens TH157, TH160, TH162, TH165, TH169, TH170, TH171, TH176, and TH186.

***Magallana belcheri* (Sowerby II, 1871)**

(Fig. 4D–F)

Ostrea belcheri Sowerby II, 1871 (in 1870–1871): pl. 7, fig. 11 (type locality: “Eastern Seas?”).

Ostrea siamensis Mörch, 1853: 62 (type locality: “Singapuhra”)—nomen nudum.

Habitat. Not recorded in the wild in the upper gulf but included here for completeness.

Material examined. Site 2, Samaesan seafood market.

Published Gulf of Thailand records. *Crassostrea belcheri* (Sowerby II, 1871): Chaitiamwong et al. (1971); Aungtonya et al. (1999); Yoosukh & Duangdee (1999); Day et al. (2000); Klinbunga et al. (2000, 2001, 2002, 2003, 2005); Yoosukh (2000); Bussarawit & Simonsen (2006a); ONEP (2007); Nabhitabhata (2009); Bussarawit & Cedhagen (2010, 2012); Phuwan et al. (2018); Trivej & Kesjinda (2018); Ninwichian et al. (2021).

Geographic range. Native to Southeast Asia, with sequenced records from Pakistan, Myanmar, India, Thailand, Malaysia, Singapore, Vietnam, China, and Indonesia (Willan et al., 2021).

Remarks. This is an important aquaculture species on both sides of the Gulf of Thailand, particularly on the eastern side (Bussarawit & Cedhagen, 2010; Trivej & Kesjinda, 2018). Three specimens purchased in the Samaesan seafood market were field identified as *Crassostrea* sp. (Table 2). *M. belcheri* is known as the white scar oyster because its white adductor muscle scar differentiates it from *M. bilineata*, the black scar oyster with its black adductor muscle scar (see below).

***Magallana bilineata* (Röding, 1798)**

(Figs. 4A–C, 6A, B)

Ostrea bilineata Röding, 1798: 170 (type locality: not stated).

Ostrea lugubris Sowerby II, 1871 (in 1870–1871): pl. 26, fig. 63 (type locality: “North America?”).

Ostrea iredalei Faustino, 1932: 546, 547, pl. 1, figs. 1–4 (type locality: “Navotas, Malabon, Parañaque, and other places on Manila Bay”, Philippines).

Ostrea madrasensis Preston, 1916: 33–35, figs. 11, 11a (type locality: “Ennur backwater, Madras”, India).

Habitat. Recorded on rocky shores and aquaculture ropes.

Material examined. Site 1, Sriracha; Site 6, mouth of Kungkraben Bay; Site 7, south Kungkraben Bay; Site 9, Pattaya. *Magallana bilineata* was also identified from the aquaculture farms in Chonburi.

Published Gulf of Thailand records. *Crassostrea bilineata* (Röding, 1798): Nabhitabhata (2009); Bussarawit & Cedhagen (2010, 2012). *Crassostrea lugubris* (Sowerby II, 1871): Aungtonya et al. (1999); Murugan et al. (1999);

Day et al. (2000); Trivej & Kesjinda (2018). *Crassostrea iredalei* Faustino, 1932: Aungtonya et al. (1999); Yoosukh & Duangdee (1999); Klinbunga et al. (2000, 2001, 2002, 2003, 2005); Yoosukh (2000); Lam & Morton (2004); Bussarawit et al. (2006); Bussarawit & Simonsen (2006a).

Geographic range. Native to Southeast Asia, with sequenced records from: India, Thailand, Malaysia, Singapore, southern Japan, and Vietnam. Unsequenced records are from Pakistan and the Philippines (Tan et al., 2024). Introduced to Queensland, Australia (Willan et al., 2021).

Remarks. *Magallana bilineata* is known as the black scar oyster because its black adductor muscle scar differentiates it from *M. belcheri*, the white scar oyster, with its white adductor muscle scar. *Magallana bilineata* is well known in Thailand where it has been variously identified as *Crassostrea bilineata*, *C. lugubris*, and *C. iredalei*. Torigoe and Bussarawit (2010) clarified that there is a single species present in Thailand (*C. bilineata*), and *C. lugubris* and *C. iredalei* are synonyms. Identified in the field as *Crassostrea* sp. (Table 2). The species was introduced to aquaculture farms in the Sriracha area about 20 years ago from Kungkraben Bay and has now become naturalised in the region (Attawut Kuntavong, pers. comm.).

Published *Magallana* records in the Gulf of Thailand rejected. *Saccostrea dactylena* (Iredale, 1939) was reported by van Gemert (2003) from Ko Samui in the southern Gulf of Thailand. *Saxostrea commercialis dactylena* Iredale, 1939 was described as an ecomorph from a specimen collected at Lindeman Island, Great Barrier Reef, Queensland. The species is accepted as valid by WoRMS (MolluscaBase eds., 2024) and is included in the genus *Magallana*. Van Gemert (2003) simply lists *S. dactylena* without a description or photograph. As there are no validated records from the Gulf of Thailand, the record is rejected.

Magallana gigas (Thunberg, 1793) was reported as *Crassostrea gigas* (Thunberg, 1793) by Aungtonya et al. (1999), Swennen et al. (2001), Robba et al. (2002), Printrakoon et al. (2008), and Sanpanich (2011). This is the Pacific oyster that has been widely introduced for aquaculture (Botta et al., 2020). It has also been reported from Singapore, but Tan et al. (2024) conducted a detailed investigation of intertidal oysters in Singapore and concluded the Singapore records of *M. gigas* were based on outdated taxonomy and the species is not present in Singapore. Similarly, the DNA sequences obtained for *Magallana* for the present paper were all *M. belcheri* and *M. bilineata*; *M. gigas* was not recorded.

Magallana rivularis (Gould, 1861) was reported as *Ostrea rivularis* Gould, 1861 by Lyngé (1909). This species is considered a taxon inquirendum by WoRMS (MolluscaBase eds., 2024). Photographs of the Lyngé specimens were obtained from the Natural History Museum of Denmark but were unidentifiable fragments.

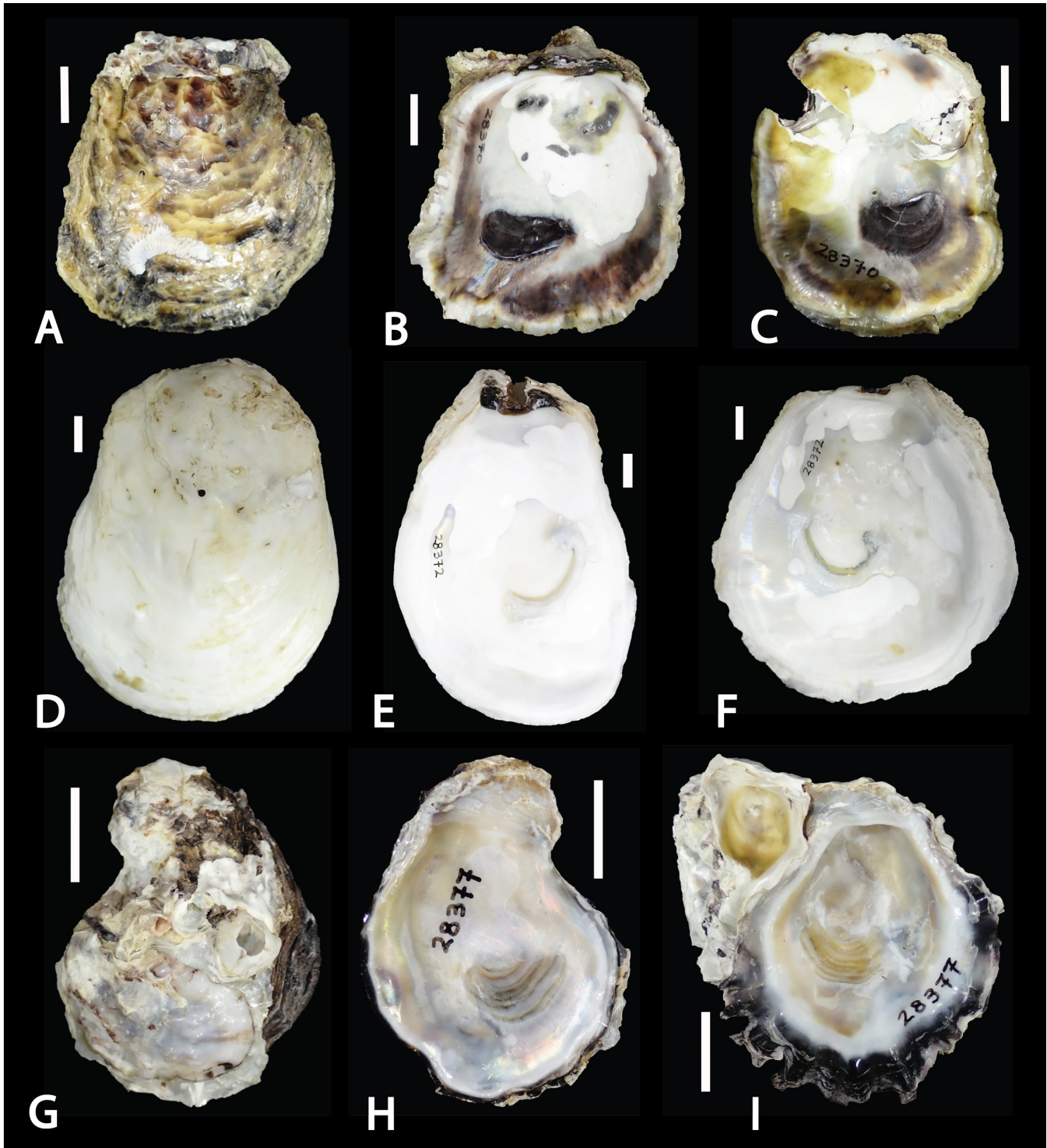


Fig. 4. *Magallana* and *Saccostrea* species collected in the upper Gulf of Thailand in August 2023. *Magallana bilineata* (A–C; ZRC.MOL.28370); *M. belcheri* (D–F; ZRC.MOL.28372); *Saccostrea scyphophilla* (G–I; ZRC.MOL.28377). For each species, the external surface of the upper (right) valve is on the left, the centre is the interior surface of the RV, and the right is the interior surface of the lower (left) valve. Scale bars are 1 cm.

Subfamily Saccostreinae Salvi & Mariottini, 2016

Genus *Saccostrea* Dollfus & Dautzenberg, 1920

Saccostrea is a widespread genus in the Indo-Pacific, with a number of species described largely on the basis of shell morphology, but *Saccostrea* taxonomy remains very confusing. Lam & Morton (2006) used genetic techniques

in an attempt to differentiate the species. They proposed a ‘superspecies’ of *S. cucullata* and a separate clade based on *S. mordax* (Gould, 1850) (= *S. scyphophilla*; see Snow et al., 2023) to include all the different ecotypes and/or forms. Recent studies indicate that 10 distinct lineages are present within *Saccostrea* (non-*mordax*); these are all probably valid species. Sekino & Yamashita (2016) argued that they should not be included in *S. cucullata*, but referred to them as

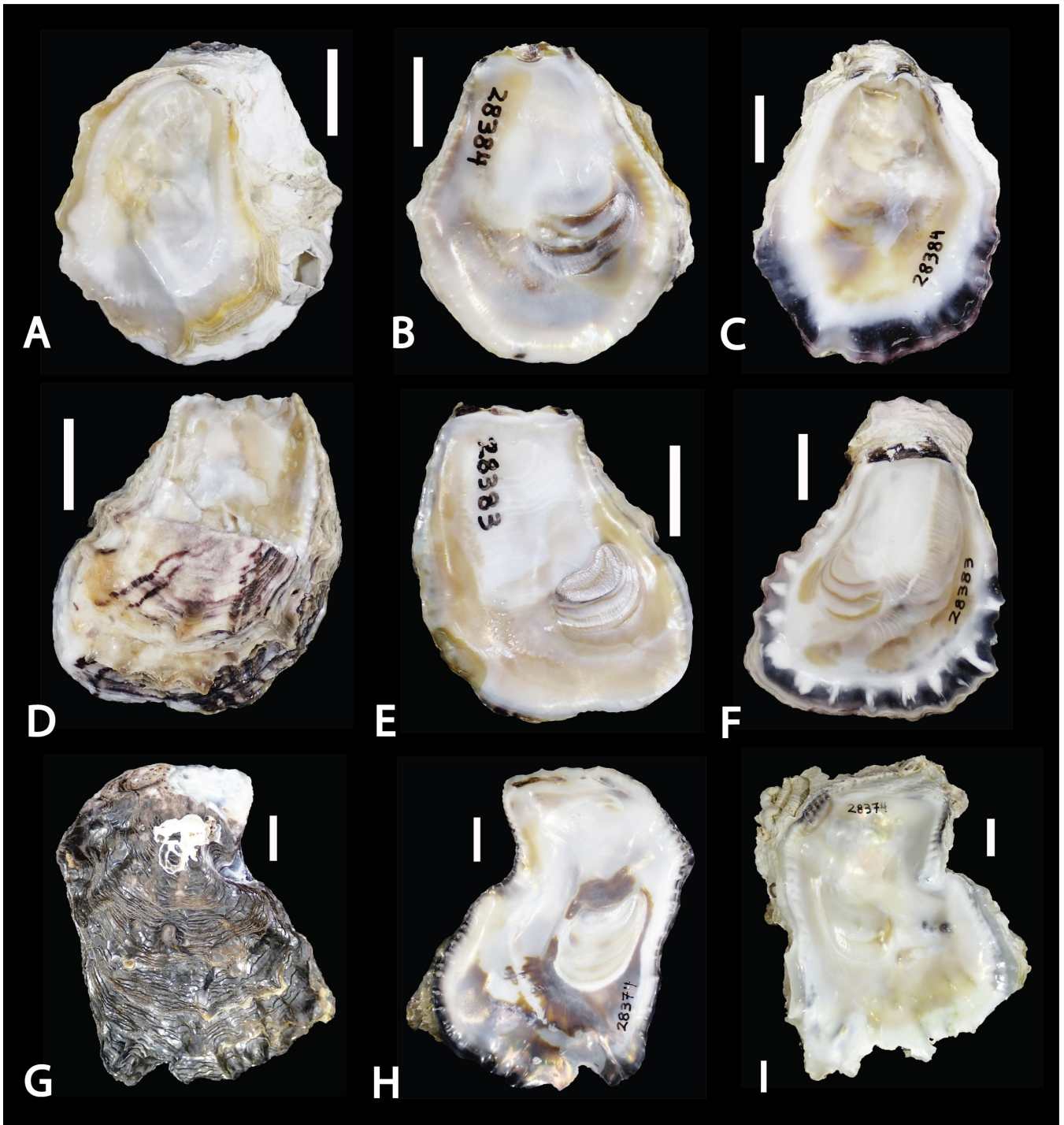


Fig. 5. *Saccostrea* species collected in the upper Gulf of Thailand in August 2023. *Saccostrea* B (A–C; ZRC.MOL.28384); *Saccostrea* F (D–F; ZRC.MOL.28383); *Saccostrea spathulata* (G–I; ZRC.MOL.28374). For each species, the external surface of the upper (right) valve is on the left, the centre is the interior surface of the RV, and the right is the interior surface of the lower (left) valve. Scale bars are 1 cm.

Saccostrea A–J. This terminology was used by Snow et al. (2023) and we have followed their recommendations. Tan et al. (in press) have recently demonstrated that *S. cucullata* was described from Ascension Island in the South Atlantic Ocean and does not occur in the Indo-West Pacific.

Visootiviset et al. (1998) pointed out the confused taxonomy of small oysters in the genus *Saccostrea* in Thailand and used a combination of electrophoretic and morphometric techniques to distinguish three species: A, B, and C. All three occurred in the Gulf of Thailand but were not named.

Klinbunga et al. (2005) also recognised three groups (1–3) of *Saccostrea* in Thailand, with groups 1 and 3 occurring in the Gulf of Thailand. These results cannot be compared with the present study as Visootiviset et al. (1998) used allozyme analysis and Klinbunga et al. (2005) used Polymerase Chain Reaction-Restriction Fragment Length Polymerase (PCR RFLP), vastly different molecular techniques from those used here.

Saccostrea species present in the upper Gulf of Thailand:

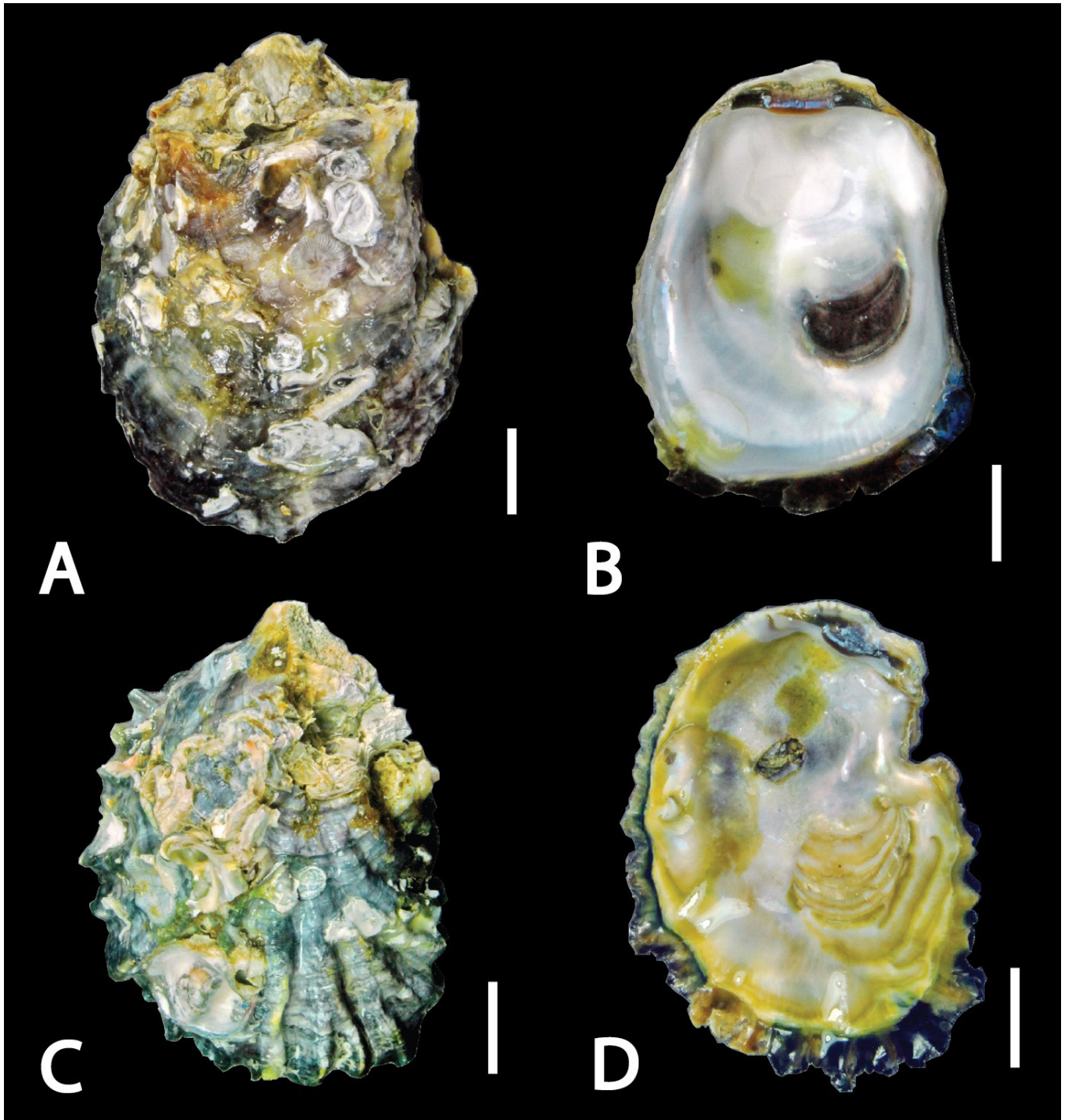


Fig. 6. Oysters from the Chonburi aquaculture farms bought in the markets in March 2024. *Magallana bilineata* (A, B; KUMF–MOLL 1333); *Saccostrea* F (C, D; KUMF–MOLL 1335). For each species, the external surface of the upper (right) valve is on the left, on the right is the interior surface of the lower (left) valve. Scale bars are 1 cm.

***Saccostrea* B species complex**
(Fig. 5A–C)

Habitat. Rocky shores, rock walls, jetty.

Material examined. Site 2, Samaesan; Site 3, Ta Kuan Beach; Site 4, Mae Rum Peung Beach; Site 5, Hin Khrong; Site 6, mouth of Kungkraben Bay; Site 7, south Kungkraben Bay; Site 9, Pattaya; Site 10, Klongwan Pier; Site 13, Kao Ta Kiep; Site 14, Cha Am. Identified in the field as

S. echinata, *S. forskahlii*, *Striostrea* sp., and *Saccostrea cuccullata* (Table 2).

Published Gulf of Thailand records. None.

Geographic range. Recorded by sequence data from Australia (Northern Territory, Queensland), China, Singapore, Vietnam, Taiwan (Lam & Morton 2006), and Kimberley, Western Australia (Snow et al., 2023).

Table 2. Summary comparison of oyster identifications in the upper Gulf of Thailand using Bussarawit & Cedhagen (2010) in the field and 16S rRNA sequencing.

DNA sequencing	Following Bussarawit & Cedhagen (2010)	Number of stations	Number sequenced
<i>Magallana belcheri</i>	<i>Crassostrea belcheri</i>	Market	3
<i>Magallana bilineata</i>	<i>Crassostrea</i> sp.	4	9
<i>Saccostrea</i> B	<i>S. cucullata</i> ; <i>S. forskahlii</i> ; <i>S. echinata</i> ; <i>Striostrea</i> sp.	10	29
<i>Saccostrea</i> F	<i>S. cucullata</i> ; <i>S. forskahlii</i> ; <i>Striostrea</i> sp.	7	15
<i>Saccostrea spathulata</i>	<i>Ostrea</i> sp.	1	7
<i>Saccostrea scyphophilla</i>	<i>S. cucullata</i>	1	4

Remarks. Published Gulf of Thailand records as *Saccostrea echinata* (Quoy & Gaimard, 1835): Aungtonya et al. (1999); Yoosukh & Duangdee (1999); Yoosukh (2000); Nabhitabhata (2009). Note: Bussarawit & Cedhagen (2010) considered Nielsen's (1976) identification of *Saccostrea cucullata* as a misidentification of *Saccostrea echinata*. WoRMS (MolluscaBase eds., 2024) considers *S. echinata* to be a valid species, but the DNA sequences identified as *S. echinata* from the upper Gulf of Thailand were *Saccostrea* B. The 16S Bayesian inference tree of Asian *Saccostrea* (Fig. 3) suggests *Saccostrea* B is paraphyletic, with more than one species taxon present. More investigation of this possibility is required, however this is outside the scope of the present paper.

***Saccostrea* F**
(Figs. 5D–F, 6C, D)

Habitat. Hanging ropes, rocky shores and pier.

Material examined. Site 1, Sriracha; Site 3, Ta Kuan Beach; Site 6, mouth of Kungkraben Bay; Site 7, south Kungkraben Bay; Site 9, Pattaya; Site 10, Klongwan Pier; Site 13, Kao Ta Kiep. Identified in the field as *Saccostrea cucullata*, *S. forskahlii*, and *Striostrea* sp. (Table 2). *Saccostrea* F was also identified from the aquaculture farms in Chonburi.

Published Gulf of Thailand records. None.

Geographic range. Taiwan and Singapore (Lam & Morton, 2006), Japan (Sekino & Yamashita, 2016), and Queensland (McDougall et al., 2024).

Remarks. Published Gulf of Thailand records as *Saccostrea forskahlii* (Bussarawit et al., 2006), specimen collected from Ang Sila Chonburi oyster farm.

***Saccostrea spathulata* (Lamarck, 1819)**
(Fig. 5G–I)

Habitat. Rocky shore.

Material examined. Site 2, Samaesan. Identified in the field as *Ostrea* sp. (Table 2).

Published Gulf of Thailand records. None.

Geographic range. Uncertain, sequenced records from Queensland, Western Australia, and Japan (McDougall et al., 2024).

Remarks. This is the species previously referred to as *Saccostrea* J. Sekino & Yamashita (2016), McDougall et al. (2024), and Richardson et al. (2024) all presented evidence to support their conclusion *Saccostrea* J is actually *S. spathulata*.

***Saccostrea scyphophilla* (Péron & Lesueur, 1807)**
(Fig. 4G–I)

Habitat. Rocky shore.

Material examined. Site 2, Samaesan. Identified in the field as *Saccostrea cucullata* (Table 2).

Published Gulf of Thailand records. Reported as *Saccostrea mordax* (Gould, 1850) by Kurozumi et al. (1989) and Yoosukh (2000).

Geographic range. Australia (Western Australia to New South Wales), Japan, China, and Taiwan (Lam & Morton, 2006). Sekino & Yamashita (2013) showed a *S. cucullata* seq (AY038076) collected from Thailand was actually *S. mordax*, but *S. mordax* is considered to be a synonym of *S. scyphophilla* (MolluscaBase eds., 2024).

Remarks. There are numerous literature references of *S. mordax* on the more open Thai coastline on the Andaman Sea, but we could only find records in the Gulf of Thailand by Kurozumi et al. (1989) and Yoosukh (2000). We detected the species only on the rocky shore at Samaesan. *Saccostrea mordax* was originally described from Fiji. While there are GenBank sequences available for the species, there are none from the type locality. Huber (2010) discusses the taxonomic history of the species in detail. *Saccostrea scyphophilla* was described by Péron & Lesueur (1807) from Shark Bay, Western Australia, and Huber (2010) considers the species to be the same, with *S. scyphophilla* having priority. Our DNA sequences of specimens collected in Samaesan (Fig. 3)

match those of Snow et al. (2023) and Wells et al. (2024), so we consider the Thai species to be *S. scyphophilla*.

Published *Saccostrea* records in the Gulf of Thailand rejected. There have been numerous reports of *S. cucullata* (Born, 1778) in the Gulf of Thailand, but unfortunately with no DNA sequences presented, it is not clear what species were actually reported. Published Gulf of Thailand records: *Ostrea cucullata* Born, 1778 reported by: Lyngø, 1909. *Saccostrea cucullata* (Born, 1778) reported by: Amornjaruchit (1988); Nateewathana (1995); Sanpanich (1998, 2011); Aungtonya et al. (1999); Yoosukh & Duangdee (1999); Klinbunga et al. (2000, 2001, 2002, 2003, 2005); Yoosukh (2000); Swennen et al. (2001); Robba et al. (2002); van Gemert (2003); Bussarawit & Simonsen (2006b); Printrakoon et al. (2008); Nabhitabhata (2009); Bussarawit & Cedhagen (2010); Samakraman et al. (2010); Negri et al. (2014).

Saccostrea commercialis (Iredale & Roughley, 1933) reported by Trivej & Kesjinda (2018) is actually *S. glomerata* (Gould, 1850), but this is a misidentification.

Saccostrea forskahlii (Gmelin, 1791). Published Gulf of Thailand records as *Saccostrea forskahlii*: Aungtonya et al. (1999); Klinbunga et al. (2000, 2001, 2005); Bussarawit et al. (2006); Bussarawit & Simonsen (2006b); Kasetsart University (2006); ONEP (2007); Nabhitabhata (2009); Bussarawit & Cedhagen (2010); Sanpanich (2011). As *S. forskahlii*: Yoosukh & Duangdee (1999). Note: Huber (2010) believes the *S. forskahlii* reported by Yoosukh & Duangdee (1999) is actually the Melanesian *S. circumscuta* (Gould, 1850). WoRMS considers *S. forskahlii* to be a synonym of *S. cucullata* (MolluscaBase eds., 2024), but it is not clear to which lineage *S. forskahlii* belongs. There are only two 16S sequences submitted to GenBank under the name *S. circumscuta*. One matches Lam & Morton (2006) lineage D and the other lineage C.

Striostrea mytiloides (Lamarck, 1819) was reported in the Gulf of Thailand by Yoosukh & Duangdee (1999) and Klinbunga et al. (2000, 2001, 2005), and *Striostrea (Parastriostrea) mytiloides* (Lamarck, 1819) by Kurozumi et al. (1989). Huber (2010) and WoRMS (MolluscaBase eds., 2024) recorded three other species of *Striostrea* Vialov, 1936: *S. denticulata* (Born, 1778) from West Africa; *S. margaritacea* (Lamarck, 1819) from South Africa and the Western Indian Ocean; and *S. prismatica* (Gray, 1825) from the Panamic Province. Huber (2010) believed *Ostrea rufa* Lamarck, 1819 from the Panamic Province may be a valid *Striostrea*, but WoRMS classifies it as *S. prismatica* (MolluscaBase eds., 2024). The Japanese *Ostrea circumscuta* Pilsbry, 1904 was included in *Striostrea* by Huber (2010) but was retained in *Ostrea* by WoRMS (MolluscaBase eds., 2024). Regardless of the final classification of these two species, the genus *Striostrea* has not been recorded from the Gulf of Thailand. *Ostrea mytiloides* Lamarck, 1819 is accepted as *Saccostrea echinata* (Quoy & Gaimard, 1835) by WoRMS (MolluscaBase eds., 2024), but there is no record of the DNA lineage of this species.

DISCUSSION

The two *Magallana* species, *M. bilineata* and *M. belcheri*, were correctly identified in the field, albeit with the earlier generic name *Crassostrea*. *Magallana bilineata* was recorded from nine sequences in the field and 16 in the markets. Field identifications of other taxa using Bussarawit & Cedhagen (2010) were inconsistent. *Saccostrea* B was identified in the field with four separate names: *S. cucullata*, *S. forskahlii*, *S. echinata*, and *Striostrea* sp. *Saccostrea* F was field identified variously as *S. cucullata*, *S. forskahlii*, and *Striostrea* sp. Both *Saccostrea* B and F were identified with more than one name on a single shoreline. *Saccostrea spathulata* and *S. scyphophilla* were found only at a single station and each was identified in the field by a single name, *Ostrea* sp. and *Saccostrea cucullata* respectively. *Saccostrea* B was the most widespread species, occurring at 10 of the 11 stations where specimens were collected, of which 29 were sequenced. *Saccostrea* F was recorded from 15 sequences in the field and 17 in the markets. *Magallana bilineata* and *Saccostrea* F occurred at all four of the markets surveyed.

There are numerous published references on the occurrence of *Saccostrea* species in the Gulf of Thailand. Unfortunately, there is no voucher material for most of these studies. Where vouchers are available, they are likely to be dry material or preserved in formalin. Visootviseth et al. (1998) described the difficulty in assessing Thai *Saccostrea* species, distinguishing three species using allozyme and morphometric techniques. Klinbunga et al. (2005) also recognised three species of *Saccostrea*, two of which occurred in the Gulf of Thailand. However, neither of these studies can be reconciled with our DNA sequences as they used different molecular techniques. The present paper reports on the upper Gulf of Thailand; further work is required to extend the results into the lower Gulf of Thailand and the Andaman coast.

Similarly, it is difficult to compare the number of species of *Saccostrea* in the upper Gulf of Thailand with other geographic areas. While numerous studies have recorded *Saccostrea* species, the lack of genetic information means the number of species reported may be different from the actual number of species in a locality. McDougall et al. (2020, 2024) found eight *Saccostrea* on a lengthy part of the Queensland coastline. Three species occur in the Kimberley of Western Australia, but only two in the Pilbara (Snow et al., 2023; Wells et al., 2024).

Oysters from aquaculture farms in Chonburi were a mixture of *M. bilineata* and *Saccostrea* F. The presence of *M. bilineata* in the aquaculture farms was to be expected, but *Saccostrea* F was unexpected as *Saccostrea* B was more widely distributed in the area and more individuals were present in the samples sequenced. Bussarawit et al. (2006) had *S. forskahlii* listed from Chonburi, which turned out to be Lineage F. While important, oyster aquaculture in the upper Gulf of Thailand is on a small scale compared to further southeast at the mouth of the Welu River on the border between Trat and Chanthaburi provinces. The key commercial species were listed as *Saccostrea commercialis*, *Crassostrea lugubris*, and

C. belcheri by Trivej & Kesjinda (2018). The taxonomy of *C. lugubris* and *C. belcheri* can be updated to *Magallana bilineata* and *M. belcheri* respectively. The *Saccostrea* is not *S. commercialis*, as this is actually the Australian temperate species *S. glomerata* (Snow et al., 2023).

The Pacific oyster *M. gigas* has been recorded on numerous occasions in the Gulf of Thailand, but these records are based on outdated concepts of oyster systematics, particularly Harry (1985). Instead, two species of *Magallana*, *M. belcheri* and *M. bilineata*, were found in the upper Gulf of Thailand. The same finding was made in a recent study of the genus in Singapore (Tan et al., 2024). Harry (1985) recognised a single species of *Crassostrea* (now *Magallana*) *gigas* in the Indo-Pacific that occurred from Pakistan to Korea and Japan, and in the Philippines, Borneo, and Sumatra. *Magallana gigas* dominates the total annual world aquaculture production of about six million tonnes (Botta et al., 2020), but this statement is based on the opinion of Harry (1985) that there is one species of *Magallana* in the Indo-Pacific. WoRMS (MolluscaBase eds., 2024) recognised 13 species of *Magallana*, five of which were described subsequent to Harry (1985). It is likely that future DNA studies will show that the aquaculture production of the genus is more widespread than just *M. gigas*.

The introduction of oysters into new parts of the world oceans, either deliberately through aquaculture or accidentally via vessels, poses a number of threats. Species may escape from the aquaculture farms and become invasive as has happened with *M. gigas* in south-eastern Australia (NIMPIS, 2022a). Other organisms, such as oyster drills, boring polychaetes, sponges, and pea crabs can be attached to or in oyster shells, or in the mantle cavity (Haupt et al., 2010). Ruesink et al. (2005) estimated that 46% of marine species introductions in northern Europe and 20% in Australia were likely to have been introduced through oyster aquaculture. Another potential impact is the introduction of diseases with introduced oysters, such as Pacific Oyster Mortality Syndrome, which can cause significant mortality in *M. gigas* (Petton et al., 2021).

The absence of *M. gigas* in Thailand demonstrated here and in Singapore (Tan et al., 2024) is important as *M. gigas* is a known invasive marine species (IMS). For example, it is one of three oysters listed by DPIRD (2016) as being of concern to Western Australia. The other two are *M. ariakensis* (Fujita in Wakita, 1929) and *Crassostrea virginica*. Because of its wide reported distribution in Southeast Asia and known invasive characteristics, *M. gigas* was the primary concern. There is direct vessel traffic between the largest port in Thailand, Laem Chabang in the study area and northwestern Australia. The absence of *M. gigas* from the upper Gulf of Thailand is thus reassuring for Australian regulators.

Two other *Magallana* (*M. bilineata* and *M. ariakensis*) have recently been recorded in Queensland and are potential IMS threats to Australia (Willan et al., 2021; NIMPIS, 2022b; Biosec Qld, 2024). *Magallana bilineata* is also a known vector for oyster pathogens (Suja et al., 2020). Thus, while there is no risk of introducing the high-risk *M. gigas* to

Australia from the upper Gulf of Thailand, there remains a lower risk of introductions from *M. bilineata* and *M. belcheri*.

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LITERATURE CITED

- Amornjaruchit S (1988) Economically important molluscan shellfish of Thailand. In: McCoy EW & Chongpeepien T (eds.) Bivalve Mollusc Culture Research in Thailand. ICLARM Technical Reports 19. Department of Fisheries, Bangkok, Thailand, International Center for Living Aquatic Resources Management, Manila, Philippines, and Deutsche Gesellschaft für Technische Zusammenarbeit (GTZ), GmbH, Eschborn, Federal Republic of Germany, pp. 1–18.
- Altschul SF, Gish W, Miller W, Myers EW & Lipman DJ (1990) Basic local alignment search tool. *Journal of Molecular Biology*, 215: 403–410.
- Aungtonya C, Thaiklang N & Srisawan D (1999) Recent records of Bivalvia in the reference collection of Phuket Marine Biological Center, Thailand. *Phuket Marine Biological Center Special Publication*, 19: 371–383.
- Backeljau T (2018) *Crassostrea gigas* or *Magallana gigas*: a community-based scientific response. *National Shellfisheries Association Quarterly Newsletter*, 2018 (1): 3.
- Bayne BL, Ahrens M, Allen SK, Angles M, Auriac D, Backeljau T, Beninger P, Bohn R, Boudry P, Davis J, Green T, Guo X, Hedgecock D, Ibarra A, Kingsley-Smith P, Krause M, Langdon C, Lapegue S, Li C, Manahan D, Mann R, Perez-Paralle L, Powell EN, Rawson PD, Speiser D, Sanchez J-L, Shumway S & Wang H (2017) The proposed dropping of the genus *Crassostrea* for all Pacific cupped oysters and its replacement by a new genus *Magallana*: A dissenting view. *Journal of Shellfish Research*, 36: 545–547.

- Bayne BL, Anglès d'Auriac M, Bacheljau T, Beninger P, Boudry P, Carnegie R, Davis J, Guo X, Hedgecock D, Krause M, Langdon C, Lapègue S, Manahani D, Mann R, Powell E & Shumway S (2019) A scientific name for Pacific oysters. *Aquaculture*, 499: 373.
- Biosec Qld (2024) Suminoe oyster (*Magallana ariakensis*) fact sheet - Aquatic invasive animals and plants - Publications | Queensland Government. <https://www.publications.qld.gov.au/dataset/aquatic-invasive-pest-animal-plant/resource/c4a3bb9f-a410-447a-a011-4bd03a716b81> (Accessed 24 April 2024).
- Born I (1778) Index rerum naturalium Musei Caesarei Vindobonensis. Pars Ima. Testacea. Verzeichniß der natürlichen Seltenheiten des k. k. Naturalien Cabinets zu Wien. Erster Theil. Schalthiere. Ex Officina Krausiana, Vindibonae, xl + 458 + 82 pp.
- Boss KJ & Jacobson MK (eds) (1985) Scarlato OA & Starobogatov YI. General evolutionary patterns and the system of the class Bivalvia [An Edited Translation]. Museum of Comparative Zoology, Harvard University, Department of Mollusks, Special Occasional Publication, 5:1–67, 9 figs.
- Botta A, Asche F, Borsuma JS & Camp EV (2020) A review of global oyster aquaculture production and consumption. *Marine Policy*, 117: 103952.
- Bussarawit S & Cedhagen T (2010) Field Guide to the Oyster Fauna of Thailand. Kyoto University Press, Japan, 47 pp.
- Bussarawit S & Cedhagen T (2012) Larvae of commercial and other oyster species in Thailand (Andaman Sea and Gulf of Thailand). *Steenstrupia*, 32: 95–162.
- Bussarawit S, Gravlund P, Glenner H & Rasmussen AR (2006) Phylogenetic analysis of Thai oyster (Ostreidae) based on partial sequences of the mitochondrial 16S rDNA gene. *Phuket Marine Biological Center Research Bulletin*, 67: 1–9.
- Bussarawit S & Simonsen V (2006a) Genetic variation in populations of white scar (*Crassostrea belcheri*) and black scar oysters (*C. iredalei*) along the coast of Thailand by means of isozymes. *Phuket Marine Biological Center Research Bulletin*, 67: 11–21.
- Bussarawit S & Simonsen V (2006b) Genetic variation in populations of four species of *Saccostrea* from Thailand, Malaysia and Australia measured by means of isozymes. *Phuket Marine Biological Center Research Bulletin*, 67: 23–37.
- Chaitiamwong S, Devahudi T & Waritswat A (1971) Review of Taxonomic Nomenclature of Some Commercially Important Shellfish in Thai Waters. Second Symposium on Marine Fisheries, April 19–20, 1971. Marine Fisheries Laboratory, Bangkok, pp. 1–28.
- Crowe TP & Frid CLJ (eds) (2015) *Marine Ecosystems: Human Impacts on Biodiversity, Functioning and Services*. Cambridge University Press, Cambridge, UK, 397 pp.
- Danic-Tchaleu G, Heurtebise S, Morga B & Lapègue S (2011) Complete mitochondrial DNA sequence of the European flat oyster *Ostrea edulis* confirms Ostreidae classification. *BMC Research Notes*, 4: 400.
- Darriba D, Taboada GL, Doallo R & Posada D (2012) jModelTest 2: more models, new heuristics and parallel computing. *Nature Methods*, 9(8): 772.
- Day AJ, Visootiviset P & Hawkins AJ (2000) Genetic diversity among cultured oysters (*Crassostrea* spp.) throughout Thailand. *Science Asia*, 26: 115–122.
- DoF (2023) Statistics of Marine Shellfish Culture. Fishery Statistics Group, Fisheries Development Policy and Planning Division, Department of Fisheries, Bangkok, Thailand. No. 10/2023, 30 pp.
- Dollfus GF & Dautzenberg P (1920) *Conchyliologie du Miocène moyen du Bassin de la Loire. Ire Partie: Pélécy-podes (suite et fin)*. Mémoires de la Société Géologique de France, Paléontologie, 22(2–4): 379–500, pls. 34–51.
- DPIRD (2016) Western Australian Prevention List for Introduced Marine Pests. Department of Primary Industry and Regional Development, Perth, WA. <http://www.fish.wa.gov.au> (Accessed 25 May 2023).
- Faustino LA (1932) Recent and fossil shells from the Philippine Islands, I. *The Philippine Journal of Science*, 49(4): 543–549.
- Fujita in Wakiya Y (1929). Japanese food oysters. *Japan Journal of Zoology*, Tokyo, 2: 359–367.
- Gmelin JF (1791) Vermes. In: Gmelin JF (ed.) *Caroli a Linnaei Systema Naturae per Regna Tria Naturae*, Ed. 13. Tome 1(6). GE Beer, Lipsiae [Leipzig]. pp. 3021–3910. *Systema Naturae*. Linnaeus (ed.). Ed. 13. 1: pars. 6.
- Gould AA (1850) Descriptions of new species of shells from the United States Exploring Expedition. *Proceedings of the Boston Society of Natural History*, 3: 151–156, 169–172, 214–218, 252–256, 275–278, 292–296, 309–312, 343–348.
- Gould AA (1861) Description of new shells collected by the United States North Pacific Exploring Expedition. *Proceedings of the Boston Society of Natural History*, 7: 385–389 [January 1861], 401–409 [February 1861]; 8: 14–32 [March 1861], 33–40 [April 1861].
- Gray JE (1825) A list and description of some species of shells not taken notice of by Lamarck. *Annals of Philosophy*, new series, 9: 407–415.
- Gastineau R, Nguyen DH, Lemieux C, Turmel M, Tremblay R, Nguyen VD, Widowati I, Witkowski A & Mouget JL (2018) The complete mitochondrial DNA of the tropical oyster *Crassostrea belcheri* from the Càn Giò mangrove in Vietnam. *Mitochondrial DNA Part B Resources*, 3: 462–463.
- Hamaguchi M, Shimabukuro H, Usuki H & Hori M (2014) Occurrences of the Indo-West Pacific rock oyster *Saccostrea cucullata* in mainland Japan. *Marine Biodiversity Records*, 7: e84.
- Hanley S (1842–1846). An illustrated and descriptive catalogue of Recent shells, by Sylvanus Hanley, B.A., F.L.S. The plates forming a third edition of the Index Testaceologicus, by William Wood. London. 272 + 18 pp., pls. 9–19. [1–224: reissue of material published in 1842–1843; 225–272, pls. 9–13: 1843; pls. 14–16: 1844; 9–18 (explanation to pls 9–13), pls. 17–19: 1846.
- Harry HW (1985) Synopsis of the supraspecific classification of living oysters (Bivalvia, Gryphaeidae and Ostreidae). *The Veliger*, 28: 121–158.
- Haupt TM, Griffiths CL, Robinson TB & Tonin AFG (2010) Oysters as vectors of marine aliens, with notes on four introduced species associated with oyster farming in South Africa. *African Zoology*, 45: 52–62.
- Herbert CJ, Humphreys J, Davies C, Roberts C, Fletcher S & Crowe T (2016) Ecological impacts of non-native Pacific oysters (*Crassostrea gigas*) and management measures for protected areas in Europe. *Biodiversity Conservation*, 25: 2835–2865.
- Huber M (2010) *Compendium of Bivalves. A Full-Color Guide to 3,300 of the World's Marine Bivalves. A Status on Bivalvia After 250 Years of Research*. Conchbooks, Hackenheim, 901 pp., CD-ROM.
- Huelsenbeck JP & Ronquist F (2001) MRBAYES: Bayesian inference of phylogenetic trees. *Bioinformatics*, 17(8): 754–755.
- Iredale T (1939) Mollusca. In: Iredale T (ed.) *Great Barrier Reef Expedition 1928–1929, Scientific Reports 5(6), Part I*. British Museum (Natural History), London, pp. 211–425.
- Iredale T & Roughley TC (1933). The scientific name of the commercial oyster of New South Wales. *Proceedings of the Linnean Society of New South Wales*, 58: 278.
- Jeamsoong S, Thaotumpitak V, Anuntawirun S, Roongrojmongkhon N & Atwill ER (2022) Meteorological and water quality factors associated with microbial diversity in coastal water from intensified oyster production areas of Thailand. *Water*, 14: 3838.
- Johnson CR & Chapman RDO (2007) Seaweed invasions: Introduction and scope. *Botanica Marina*, 50: 321–325.

- Kasetsart University (2006) Biodiversity Survey and Information System Project: Final Report. Volume 3. Marine and Coastal Biodiversity and Ecology. Forest Research Center, Faculty of Forestry, Kasetsart University, Bangkok, pp. 11–117.
- Katsanevakis S, Wallentinus I, Zenetos A, Leppäkoski E, Çinar ME, Öztürk B, Grabowski M, Golani D & Cardoso AC (2014) Impacts of invasive alien marine species on ecosystem services and biodiversity: a pan-European review. *Aquatic Invasions*, 9: 391–423.
- Klinbunga S, Ampayup P & Tassanakajon A (2001) Genetic diversity and molecular markers of cupped oysters (Genera *Crassostrea*, *Saccostrea*, and *Striostrea*) in Thailand revealed by randomly amplified polymorphic DNA analysis. *Marine Biotechnology*, 3: 133–144.
- Klinbunga S, Ampayup P, Tassanakajon A, Jarayabhand P & Yoosukh W (2000) Development of species-specific markers of the tropical oyster (*Crassostrea belcheri*) in Thailand. *Marine Biotechnology* 2: 476–484.
- Klinbunga S, Ampayup P, Khamnamtong N, Tassanakajon A, Jarayabhand P & Yoosukh W (2002) Molecular genetic markers for taxonomy of oysters in Thailand. *Fisheries Science, Supplement 2*, 68: 1087–1090.
- Klinbunga S, Khamnamtong B, Puanglarp N, Jarayabhand P, Yoosukh W & Menasveta P (2005) Molecular taxonomy of cupped oysters (*Crassostrea*, *Saccostrea*, and *Striostrea*) in Thailand based on COI, 16S, and 18S rDNA polymorphism. *Marine Biotechnology*, 7: 306–317.
- Klinbunga S, Khamnamtong N, Tassanakajon A, Puanglarp N, Jarayabhand P & Yoosukh W (2003) Molecular genetic identification tools for three commercially cultured oysters (*Crassostrea belcheri*, *Crassostrea iredalei*, and *Saccostrea cucullata*) in Thailand. *Marine Biotechnology*, 5: 27–36.
- Kurozumi T, Kosuge T & Tsuchiya M (1989) List of shallow-water marine molluscs in the Sichang Island, the Gulf of Thailand. *Galaxea*, 8: 295–310.
- Lam K & Morton B (2004) The oysters of Hong Kong (Bivalvia: Ostreidae and Gryphaeidae). *The Raffles Bulletin of Zoology*, 52: 11–28.
- Lam K & Morton B (2006) Morphological and mitochondrial-DNA analysis of the Indo-West Pacific rock oysters (Ostreidae: *Saccostrea* species). *Journal of Molluscan Studies*, 72(3): 235–245.
- Lamarck JBPA (1819) Histoire naturelle des animaux sans vertèbres, présentant les caractères généraux et particuliers de ces animaux, leur distribution, leurs classes, leurs familles, leurs genres, et la citation des principales espèces qui s'y rapportent; précédée d'une Introduction offrant la détermination des caractères essentiels de l'Animal, sa distinction du végétal et des autres corps naturels; enfin, l'exposition des principes fondamentaux de la Zoologie. Tome sixième. Ire. partie. L'auteur, Paris, 343 pp.
- Li XX & Qi ZY (1994) Studies on the comparative anatomy, systematic classification and evolution of Chinese oysters. *Studia Marina Sinica*, 35: 143–173. [In Chinese]
- Li C, Haws M, Wang H & Guo X (2017) Taxonomic classification of three oyster (Ostreidae) species from Myanmar. *Journal of Shellfish Research*, 36: 365–371.
- Linnaeus C (1758) *Systema naturae per regna tria naturae, secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis, locis*. Tomus I. Editio decima, reformata. Laurentii Salvii, Holmiae, 823 pp.
- Liu S, Liu Y, He J, Lin Z & Xue Q (2022) The complete mitochondrial genome of *Crassostrea hongkongensis* from East China Sea indicates species' range may extend northward. *Molecular Biology Reports*, 49: 1631–1635.
- Lynge H (1909) *Marine Lamellibranchia*. The Danish Expedition to Siam 1899–1900. Det Kongelige Danske Videnskabernes Selskab Skrifter, Naturvidenskab og Mathematisk Afdeling, 5: 99–299.
- McDonald JI, Wellington CM, Coupland GT, Pedersen D, Kitchen B, Bridgwood SD, Hewitt M, Duggan R & Abdo DA (2020) A united front against marine invaders: Developing a cost-effective marine biosecurity surveillance partnership between government and industry. *Journal of Applied Ecology*, 57: 77–84.
- McDougall C, Nenadic N & Healy J (2020) *Guide to Queensland's Intertidal Oysters*. Griffith University, Brisbane, Queensland, Australia. 16 pp.
- McDougall C, Nenadic N, Richardson M & Healy JM (2024) Molecular identification of intertidal rock oyster species in north-eastern Australia reveals new candidates for aquaculture. *Aquaculture*, 587: 740838.
- MolluscaBase eds. (2024). MolluscaBase. World Register of Marine Species (WoRMS). <https://www.marinespecies.org/aphia.php?p=taxdetails&id=140656> (Accessed 8 December 2024).
- Molnar JL, Gamboa RL, Revenga C & Spalding MD (2008) Assessing the global threat of invasive species to marine biodiversity. *Frontiers in Ecology and the Environment*, 6: 485–492.
- Mörch OAL (1853) *Catalogus conchyliorum quae reliquit D. Alphonso d'Aguirra et Gadea Comes de Yoldi, Regis Daniae Cubiculariorum Princeps, Ordinis Dannebrogici in Prima Classe & Ordinis Caroli Tertii Eques. Fasciculus Secundus. Acephala. Annulata Cirripedia. Echinodermata. Haec conchylia publica auctione calendis Juniiis dividuntur*. Ludovici Klein, Hafniae, ii + 74 + 2 pp.
- Murugan A, Niklasson M, Bussarawit S, Aungtonya C & Boneka F (1999) Allozyme comparisons of black-scar oyster populations of India, Thailand, and Indonesia. *Phuket Marine Biological Center Special Publication*, 19: 139–144.
- Nabhitabhata J (2009) *Checklist of Mollusca Fauna in Thailand*. Office of Natural Resources and Environmental Policy and Planning, Bangkok, 576 pp.
- Nateewathana A (1995) Taxonomic account of commercial and edible molluscs, excluding cephalopods, of Thailand. *Phuket Marine Biological Center Special Publication*, 15: 93–116.
- Negri MP, Sanfilippo R, Basso D, Rosso A & Geronimo SID (2014) Molluscan associations from the Pak Phanang Bay (SW Gulf of Thailand) as a record of natural and anthropogenic changes. *Continental Shelf Research*, 84: 204–218.
- Nielsen C (1976) An illustrated checklist of bivalves from PMBC beach with a reef-flat at Phuket. *Phuket Marine Biological Center Research Bulletin*, 9: 1–7.
- NIMPCG [National Introduced Marine Pests Coordinating Group] (2010a) *Marine pests monitoring manual: Version 2*. National Introduced Marine Pests Coordinating Group, Department of Agriculture, Fisheries and Forestry, Canberra, Australia.
- NIMPCG [National Introduced Marine Pests Coordinating Group] (2010b) *Australian marine pests monitoring guidelines: Version 2*. National Introduced Marine Pests Coordinating Group, Department of Agriculture, Fisheries and Forestry, Canberra, Australia.
- NIMPIS (2022a) *Species – Magallana (Crassostrea) gigas*. National Introduced Marine Pests Identification System, Canberra ACT. <https://nimpis.marinepests.gov.au/species/species/133> (Accessed 17 June 2022).
- NIMPIS (2022b) *Species - Magallana (Crassostrea) bilineata*. National Introduced Marine Pests Identification System, Canberra ACT. <https://nimpis.marinepests.gov.au/species/species/151> (Accessed 17 June 2022).
- Ninwichian P, Ruangsri J, Phuwan N, Khamnamtong B & Klinbunga S (2021) Development of polymorphic microsatellites for genetic studies of white scar oyster (*Crassostrea belcheri*) using paired-end shotgun sequencing. *Molecular Biology Reports*, 48: 4273–4283.

- Ó'Foighil D, Gaffney PM & Hilbish TJ (1995). Differences in mitochondrial 16S ribosomal gene sequences allow discrimination among American [*Crassostrea virginica* (Gmelin)] and Asian [*C. gigas* (Thunberg), *C. ariakensis* Wakiya] oyster species. *Journal of Experimental Marine Biology and Ecology*, 192: 211–220.
- ONEP (2007) Final Report: Biodiversity Survey and Information System of the Biodiversity Hotspots; Petburi and Prachuap Khiri Khan Provinces. Office of Natural Resources and Environmental Policy and Planning, Bangkok, 1310 pp.
- Péron F & Lesueur CA in Péron F (1807) Voyage de découvertes aux terres Australes, exécuté par Ordre de Sa Majesté l'Empereur et Roi, sur les Corvettes le Géographe, le Naturaliste, et la Goëlette le Casuarina, pendant les années 1800, 1801, 1802, 1803 et 1804; publié par décret impérial, sous le Ministère de M. de Champagny. Tome premier. Historique. Imprimerie Impériale, Paris, xv + 496 + 2 + xv pp.
- Petton B, Destoumieux-Garzó D, Pernet F, Toulza E, de Lorgeril J, Degremont L & Mitta G (2021) The Pacific Oyster Mortality Syndrome, a polymicrobial and multifactorial disease: state of knowledge and future directions. *Frontiers in Immunology*, 12: 630343.
- Phuwan N, Ninwichian P, Khemklad S & Khamnamtong B (2018) Development of polymorphic microsatellites in white scar oyster *Crassostrea belcheri*. *Chiang Mai Journal of Science*, 45: 2666–2678.
- Pilsbry HA (1904). New Japanese marine Mollusca: Pelecypoda. *Proceedings of the Academy of Natural Sciences of Philadelphia*, 56: 550–561, pls. 39–41.
- Preston HB (1916) Report on a collection of Mollusca from the Cochin and Ennur backwaters. *Records of the Indian Museum*, 12: 27–39.
- Printrakoon C, Wells FE & Chitramvong Y (2008) Distribution of molluscs in mangroves at six sites in the upper Gulf of Thailand. In: Bieler R, Chalermwat K, Mikkelsen P, Tan KS & Wells FE (eds.) *Molluscs of Eastern Thailand*. *Proceedings of the International Marine Bivalve Workshop, Chanthaburi, Thailand, August–September 2005, with contributions on other molluscan groups*. *Raffles Bulletin of Zoology, Supplement* 18: 247–257.
- Quoy JRC & Gaimard JP (1832–1835) Voyage de la corvette l'Astrolabe: exécuté par ordre du roi, pendant les années 1826–1827–1828–1829, sous le commandement de M. J. Dumont d'Urville. *Zoologie*. 1: i–I, 1–264; 2(1): 1–321 [1832]; 2(2): 321–686 [1833]; 3(1): 1–366 [1834]; 3(2): 367–954 [1835]; *Atlas (Mollusques)*: pls 1–93 [1833].
- Rafinesque CS (1815) *Analyse de la nature ou tableau de l'univers et des corps organisés. Le nature es mon guide, et Linnéus mon maître*. Privately published, Palermo, 224 pp.
- Rambaut A, Drummond AJ, Xie D, Baele G & Suchard MA (2018) Posterior summation in Bayesian phylogenetics using tracer 1.7. *Systematic Biology*, 67: 901–904.
- Ren J, Hou Z, Wang H, Sun MA, Liu X, Liu B & Guo X (2016) Intraspecific variation in mitogenomes of five *Crassostrea* species provides insight into oyster diversification and speciation. *Marine Biotechnology*, 18: 242–254.
- Richardson MA, Nenadic N, Wingfield M & McDougall C (2024) The development of multiplex PCR assays for the rapid identification of multiple *Saccostrea* species, and their practical applications in restoration and aquaculture. *BMC Ecology and Evolution*, 24: 67.
- Robba E, Di Geronimo I, Chaimanee N, Negri M & Sanfilippo R (2002) Holocene and recent shallow soft-bottom mollusks from the northern Gulf of Thailand area: Bivalvia. *Bollettino Malacologico*, 38: 49–132.
- Röding PF (1798) *Museum Boltenianum sive catalogus cimeliorum e tribus regnis naturae. Pars secunda continens conchyliam sive testacea univalvia, bivalvia & multivalvia*. Johan. Christi. Trappii, Hamburgi. 199 pp.
- Ruesink JL, Lenihan HS, Trimble AC, Heiman KW, Micheli F, Byers JE & Kay MC (2005) Introduction of non-native oysters: ecosystem effects and restoration implications. *Annual Review of Ecology, Evolution and Systematics*, 36: 643–689.
- Sacco F (1897) *I Molluschi dei terreni terziarii del Piemonte e della Liguria, Parte 23 (Ostreidae, Anomiidae, Dimyidae)*. *Bollettino dei Musei di Zoologia ed Anatomia Comparata della R. Università di Torino*. 12(298): 99–100.
- Salimi PA, Creed JC, Esch MM, Fenner D, Jaafar Z, Levesque JC, Montgomery AD, Salimi MA, Edward JKP, Raj KD & Sweet M (2021) A review of the diversity and impact of invasive non-native species in tropical marine ecosystems. *Marine Biodiversity Records*, 14: 11.
- Salvi D, Macali A & Mariottini P (2014) Molecular phylogenetics and systematics of the bivalve family Ostreidae based on rRNA sequence-structure models and multilocus species tree. *PLoS ONE*, 9(9): e108696.
- Salvi D & Mariottini P (2016) Molecular taxonomy in 2D: a novel ITS2 rRNA sequence-structure approach guides the description of the oysters' subfamily Saccostreinae and the genus *Magallana* (Bivalvia: Ostreidae). *Zoological Journal of the Linnean Society*, 179: 263–276.
- Salvi D & Mariottini P (2021) Revision shock in Pacific oyster taxonomy: the genus *Magallana* (formerly *Crassostrea* in part) is well-founded and necessary. *Zoological Journal of the Linnean Society*, 192: 43–58.
- Salvi D, Bertsch H, Caceres-Martínez J, Cruz Flores R, Del Rio-Portilla MA, Eernisse D, Healy JM, Lafarga-De La Cruz F, Londono-Cruz E, McDougall C, Oliver GP, Oliverio M, Paniaguac C, Willan RC, Zacherl DC & Mariottini P (2022) Taxonomic discussion on scientific names for Pacific oysters requires evidence-based arguments and pluralism. *Aquaculture*, 546: 737298.
- Samakraman S, Williams GA & Ganmanee M (2010) Spatial and temporal variability of intertidal rocky shore bivalves and gastropods in Sichang Island, east coast of Thailand. *Publications of the Seto Marine Biological Laboratory, Special Publication Series*, 10: 35–46.
- Sanpanich K (1998) An annotated check list of marine bivalves from Chonburi and Rayong Province, the east coast of Thailand. *Phuket Marine Biological Center Special Publication*, 18: 297–306.
- Sanpanich K (2011) Marine bivalves occurring on the east coast of the Gulf of Thailand. *ScienceAsia*, 37: 195–204.
- Scarlato OA & Starobogatov YI (1979) Osnovy cherty evoliutsii i sistema klassa Bivalvia [General evolutionary patterns and the system of the Class Bivalvia]. In: Starobogatov YI (ed.) *Morfologiya, Sistematika i Filogeniya Molliuskov [Morphology, Systematics and Phylogeny of Mollusks]*. *Akademiia Nauk SSSR, Trudy Zoologicheskogo Instituta*, 80: 5–38. [In Russian. For English translation, see Boss KJ & Jacobson MK (1985)].
- Sekino M & Yamashita H (2013) Mitochondrial DNA barcoding for Okinawan oysters: a cryptic population of the Portuguese oyster *Crassostrea angulata* in Japanese waters. *Fisheries Science*, 79: 61–76.
- Sekino M, Ishikawa H, Fujiwara A, Doyola-Solis EFC, Lebata-Ramos MJH & Yamashita H (2015) The first record of a cupped oyster species *Crassostrea dianbaiensis* in the waters of Japan. *Fisheries Science*, 81: 267–281.
- Sekino M & Yamashita H (2016) Mitochondrial and nuclear DNA analyses of *Saccostrea* oysters in Japan highlight the confused taxonomy of the genus. *Journal of Molluscan Studies*, 82: 492–506.
- Sigwart JD, Wong NLWS & Esa Y (2021) Global controversy in oyster systematics and a newly described species from SE Asia

- (Bivalvia: Ostreidae: Crassostreinae). *Marine Biodiversity*, 51: 83.
- Simon C, Frati F, Beckenbach A, Crespi B, Liu H & Flook P (1994) Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Annals of the Entomological Society of America*, 87: 651–701.
- Snow M, Fotedar S, Wilson NG & Kirkendale LA (2023) Clarifying the natural distribution of *Saccostrea* Dollfus and Dautzenberg, 1920 (edible rock oyster) species in Western Australia to guide development of a fledgling aquaculture industry. *Aquaculture*, 566: 739202.
- Sowerby GB II (1870–1871) Monograph of the genus *Ostraea*. *Conchologia Iconica*, 18: unpaginated text [plate captions], pls. 1–33.
- Suja G, Lijo J, Kripa V, Sunil Mohamed K & Sanil NK (2020) A comparison of parasites, pathological conditions and condition index of wild and farmed populations of *Magallana bilineata* (Roding, 1798) from Vembanad Lake, west coast of India. *Aquaculture*, 515: 734548.
- Swennen C, Moolenbeek RG, Ruttanadukul N, Hobbelink H, Dekker H & Hajisamae S (2001) The molluscs of the southern Gulf of Thailand. *Thai Studies in Biodiversity*, 4: 1–210.
- Szuster BW, Chalermwat K, Flaherty M & Intacharoen P (2008) Peri-urban oyster farming in the upper Gulf of Thailand. *Aquaculture Economics & Management*, 12: 268–288.
- Tan KS, Tan SK, Lukehurst SS & Wells FE (2024) Assessing the threat of the oyster genus *Magallana* (Bivalvia: Ostreidae) in Singapore to the Australian marine environment. *Raffles Bulletin of Zoology*, 72: 162–183.
- Tan SK, Wells FE, Tan KS, Lukehurst S, Morgan M & Fotedar S (in press) Identity of the enigmatic oyster *Saccostrea cucullata* (Bivalvia: Ostreidae). *Journal of Molluscan Studies*.
- Tantanasiriwong R (1979) A checklist of marine bivalves from Phuket Island, adjacent mainland and offshore islands, western peninsular Thailand. *Phuket Marine Biological Center Research Bulletin*, 27: 15.
- Thunberg CP (1793) Tekning och Beskrifning på en stor Ostronsort ifrån Japan. *Kongliga Vetenskaps Academiens Nya Handlingar*, 14(4–6): 140–142, pl. 6.
- Torigoe K & Bussarawit S (2010) Validation of the commercial cultured black scar oyster species name, *Crassostrea bilineata* rather than *Crassostrea lugubris* and *Crassostrea iredalei* in Indo-Pacific region. Chulalongkorn University, Bangkok, Thailand. *Tropical Natural History, Supplement 3*: 235.
- Trivej P & Kesjinda K (2018) The onset and withdrawal of the rainy season in Thailand and their effects on oyster farming. *Environmental Development*, 27: 118–112.
- van Gemert L (2003) Marine schelpen verzamelen op Ko Samui (Golf van Thailand). *De Kreukel*, 39: 95–109.
- Vessel-Check (2022) Vessel-Check Aquatic Biosecurity Solutions. <https://vessel-check.com/> (Accessed 17 November 2022).
- Vialov O (1936) Sur la classification des huîtres. *Comptes Rendus (Doklady) de l'Académie des Sciences de l'URSS. Ser. 2*, 4(1): 17–20.
- Visootviseth P, Day A & Siwadune T (1998) Electrophoretic and morphometric analyses in species differentiation of small oysters, *Saccostrea* spp., in Thailand. *Journal of the Science Society of Thailand*, 24: 24–26.
- Wang H, Guo X, Zhang G & Zhang F (2004a) Classification of Jinjiang oysters *Crassostrea rivularis* (Gould, 1861) from China, based on morphology and phylogenetic analysis. *Aquaculture*, 242: 137–155.
- Wang Y, Xu Z & Guo X (2004b) Differences in the rDNA-bearing chromosome divide the Asian-Pacific and Atlantic species of *Crassostrea* (Bivalvia, Mollusca). *Biological Bulletin*, 206: 46–54.
- Wells FE, Lukehurst SS, Fullwood LAF & Harvey ES (2024) Distribution of intertidal rock oysters in the Pilbara, Western Australia. *Management of Biological Invasions*, 15: 131–143.
- Willan RC (2021) *Magallana* or mayhem? *Molluscan Research*, 41: 75–79.
- Willan RC, Nenadic N, Ramage A & McDougall C (2021) Detection and identification of the large, exotic, crassostreine oyster *Magallana bilineata* (Röding, 1798) in northern Queensland, Australia. *Molluscan Research*, 41: 64–74.
- Yoosukh W & Duangdee T (1999) Living oyster in Thailand. *Phuket Marine Biological Center Special Publication*, 19: 363–370.
- Yoosukh W (2000) Taxonomic account of oysters in Thailand. *Mollusk Research in Asia, Proceedings of the Special Session on Mollusk Research in Asia held at 5th Asian Fisheries Forum, on November 12, 1998*. Chulalongkorn University, Bangkok, Thailand, pp. 81–86.
- Yu Z, Wei Z, Kong X & Shi W (2008) Complete mitochondrial DNA sequence of oyster *Crassostrea hongkongensis*—a case of “Tandem duplication-random loss” for genome rearrangement in *Crassostrea*? *BMC Genomics*, 9: 477.

Appendix 1. List of 16S sequences retrieved from GenBank and utilised in the Bayesian Inference analysis, in addition to those provided in Table 1 (main text).

Species	GenBank accession number	Location	References
<i>Ostrea edulis</i>	JF274008	France	Danic-Tchaleu et al. (2011)
<i>Magallana bilineata</i>	MT985154	India	N/A
	MT579758	Australia: Cairns	Willan et al. (2021)
	AB972008	Japan: Okinawa, Nago, Haneji-naikai	Sekino et al. (2015)
<i>Magallana belcheri</i>	KX961675	Myanmar	Li et al. (2017)
	MH051332	Vietnam	Gastineau et al. (2018)
	AY160758	China: Hong Kong	Lam & Morton (2004)
<i>Magallana gigas</i>	MZ400416	Australia: Port Adelaide, South Australia	N/A
	KJ855245	China: Haihua, Weifang, Shandong	Ren et al. (2016)
	KJ855241	Japan: Komaru	Ren et al. (2016)
<i>Magallana dianbaiensis</i>	LC120776	Japan: Wakayama, Shirahama, Tonda-River	N/A
	LC120775	Japan: Wakayama, Tanabe, Aidzu-River	N/A
	AB971988	Japan: Ehime, Minami-uwa, Misho Bay	Sekino et al. (2015)
<i>Magallana hongkongensis</i>	EU266073	China	Yu et al. (2008)
	MZ337404	China: Sanmen Bay, East China Sea	Lui et al. (2022)
	AY632549	China: Beihai	Wang et al. (2004b)
<i>Saccostrea</i> lineage A	AY247371	Australia: Carnarvon, Western Australia	Lam & Morton (2006)
	MN153602	Australia: Dampier, Western Australia	Snow et al. (2023)
	MN153698	Australia: Shark Bay, Western Australia	Snow et al. (2023)
	MT124270	Australia: Western Australia	Snow et al. (2023)
<i>Saccostrea</i> lineage B	MT124282	Australia: Kimberley, Western Australia	Snow et al. (2023)
	OR340024	Australia: Queensland	McDougall et al. (2024)
	AY247336	China: Cape D'Aguilar, Hong Kong	Lam & Morton (2006)
	AY247392	China: Sanya	Lam & Morton (2006)
<i>Saccostrea</i> lineage C	AY247380	Japan: Okinawa	Lam & Morton (2006)
	LC111198	Japan: Kagoshima, Amami-Oshima Island	Sekino & Yamashita (2016)
	LC111156	Japan: Kagoshima, Amami-Oshima Island	Sekino & Yamashita (2016)
<i>Saccostrea</i> lineage D	AY247321	Taiwan: Shiman	Lam & Morton (2006)
	AY247391	China: Sanya	Lam & Morton (2006)
<i>Saccostrea</i> lineage E	AY247328	Australia: Darwin Harbour, Northern Territory	Lam & Morton (2006)
	AY247387	Taiwan: Shiman	Lam & Morton (2006)
	LC268852	Japan: Kochi	N/A
<i>Saccostrea</i> lineage F	AY247384	Taiwan: Shiman	Lam & Morton (2006)
	LC111254	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
	LC111219	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
<i>Saccostrea</i> lineage G	AY247385	Taiwan: Shiman	Lam & Morton (2006)
	LC111202	Japan: Kagoshima, Amami-Oshima Island	Sekino & Yamashita (2016)
	LC155011	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
	OR340012	Australia: Percy Islands	McDougall et al. (2024)
<i>Saccostrea</i> lineage H	LC111262	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
	LC155015	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
<i>Saccostrea</i> lineage I	LC111211	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
	LC111214	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
	LC111215	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
	OR340007	Australia: Orpheus Island	McDougall et al. (2024)

Species	GenBank accession number	Location	References
<i>Saccostrea</i> lineage J	LC111222	Japan: Okinawa, Iriomote Island, Yaeyama	Sekino & Yamashita (2016)
	LC111260	Japan: Okinawa, Iriomote Islnd, Yaeyama	Sekino & Yamashita (2016)
	OR340003	Australia: Orpheus Island	McDougall et al. (2024)
	MN153593	Australia: Cone Bay, Kimberley, Western Australia	Snow et al. (2023)
<i>Saccostrea</i> <i>kegaki</i>	AY247317	Japan: Morozaki	Lam & Morton (2006)
	LC111090	Japan: Kagoshima, Kakeroma Island	Sekino & Yamashita (2016)
<i>Saccostrea</i> <i>glomerata</i>	AY247341	Australia: Albany, Western Australia	Lam & Morton (2006)
	AY247288	Australia: La Parouse, Sydney, New South Wales	Lam & Morton (2006)
	MN153625	Australia: Oyster Harbour, Albany, Western Australia	Snow et al. (2023)
	OR340034	Australia: Round Island Urangan	McDougall et al. (2024)
<i>Saccostrea</i> <i>scyphophilla</i>	AY247363	Australia: Quobba, Western Australia	Lam & Morton (2006)
	MN153617	Australia: South of Onslow, Western Australia	Snow et al. (2023)
	MN450281	Australia: Kalbarri, Western Australia	Snow et al. (2023)
	OR339998	Australia: Minjerrabah, Queensland	McDougall et al. (2024)
	AY247339	Australia: Cooe Bay, Rockhampton, Queensland	Lam & Morton (2006)
	MN450285	Australia: Kalbarri, Western Australia	Snow et al. (2023)
<i>Saccostrea</i> <i>mordoides</i>	OR339997	Australia: Hook Island	McDougall et al. (2024)
	OR339996	Australia: Orpheus Island	McDougall et al. (2024)
	AB898224	Japan: Okinawa, Ishigaki Island	Hamaguchi et al. (2014)
	AB898223	Japan: Okinawa, Ishigaki Island	Hamaguchi et al. (2014)